

SLOW DECLINE OF BLACK PEPPER (*PIPER NIGRUM L.*) AND ROLE OF PLANT PARASITIC NEMATODES AND *PHYTOPHTHORA CAPSICI* IN THE DISEASE COMPLEX

K.V. RAMANA, Y.R. SARMA and C. MOHANDAS¹

National Research Centre for Spices, Calicut 673 012, Kerala

ABSTRACT

A field trial on management of slow decline of black pepper (*Piper nigrum L.*) in arecanut-black pepper mixed cropping system was taken up with soil application of phorate, neem cake and bavistin. Application of nematicide and neem cake was highly effective in reducing the populations of *Meloidogyne incognita* and *Radopholus similis*. However, treatments did not show clear evidence of remission of slow decline symptoms. Moreover, during the course of three years of trial the vine death ranged from 16.6 to 58.3 per cent in different treatments. *Phytophthora capsici* was isolated from feeder roots of disease affected vines. This suggests that *P. capsici* also may be playing a role in the slow decline complex. High percentage of mortality in bavistin treated vines is another indirect supporting evidence for *Phytophthora* involvement in root rot since this fungus is insensitive to bavistin. There appears to be a lack of spatial segregation of plant parasitic nematodes and *Phytophthora* under field conditions. The studies strongly indicate the need for integrated disease management to check both plant parasitic nematodes and *Phytophthora* and to boost the vigour and productivity of the pepper vines.

INTRODUCTION

Slow decline (slow wilt) and *Phytophthora* foot rot (quick wilt) of black pepper (*Piper nigrum L.*) are the major constraints in pepper production. These diseases are prevalent in all the pepper growing tracts in Kerala and Karnataka. Plant parasitic nematodes viz., *Meloidogyne incognita* and *Radopholus similis* were reported to be primarily responsible for slow decline (van der Vecht, 1950; Christie, 1957; Ichinoh, 1975 & 1976; Ting, 1975; Venkitesan and Setty, 1977; Mustika, 1978 and Ramana et al., 1987). Pathogenicity tests conducted under simulated field conditions at National Research Centre for Spices, Calicut, Kerala, have shown that both the nematode species caused significant damage to root system leading to leaflet yellowing, defoliation and die back symptoms which are typical of slow decline syndrome (Mohandas & Ramana, Unpublished). In a pure plantation, application of phorate 10G @ 3 g a.i./vine, twice in a year significantly reduced the nematode population and improved the health of the vines (Anonymous, 1985). According to Hubert (1957) and Bridge (1978), *R. similis* was primarily responsible for the disease in Indonesia,

but an association with fungus like *Fusarium* sp. was necessary to cause 'Yellows' disease. Namibiar and Sarma (1979) opined that the disease is complex in nature involving nematode-fungal complex coupled with nutrient deficiency and soil moisture stress. Consistent association of *Fusarium* sp. with the roots of slow decline affected vines was reported (Namibiar and Sarma, 1977) though the role of the fungus in the disease was not understood then. In view of the known efficacy of bavistin in the control of *Fusarium* sp. and in order to get indirect information on the role of *Fusarium*, if any, field trial for disease management was undertaken to study the efficacy of phorate alone and in combination with bavistin in reducing the disease incidence in areca-pepper cropping system.

MATERIALS AND METHODS

The trial was conducted in an arecanut based black pepper plantation at Central Plantation Crops Research Institute, Regional Station, Vittal (Karnataka) during 1985-86. A total of 216 pepper vines in a contiguous block were selected for the experiment with the following treatments. The experiment was

¹Present Address: P. O. Box No. 100, Kozhikode, Kerala.

replicated thrice with a plot size of 12 vines. Recommended package of practices were adopted throughout the trial. Treatments : T_1 - Control (No treatment), T_2 - Phorate 10 G @ 1.5 g a.i./vine, T_3 - Phorate 10 G @ 3 g a.i./vine, T_4 - Bavistin 0.2% soil drench @ 5 l/vine, T_5 - Phorate 10 G @ 3 g a.i./vine + Bavistin 0.2% soil drench @ 5 l/vine and T_6 - Neem cake @ 2 kg/vine.

The treatments were given twice a year, first application during May/June and second application during September/October. Root samples were collected every year during September/October prior to second application of treatments. Populations of *M. incognita* and *R. similis* in the roots were estimated. The roots were also tested for the presence of *Phytophthora* using a selective medium PVPH. Disease symptoms such as foliar yellowing was recorded every year on the basis of visual scoring on 0-3 (0=Healthy, 1=Mild foliar yellowing, 2=Medium foliar yellowing, and 3=Severe foliar yellowing).

RESULTS AND DISCUSSION

Population of plant parasitic nematodes viz.,

Table I. Populations of root knot nematode, *Meloidogyne incognita* in black pepper (per 5 g root)

Treatment	1985 (pi)	1986	1987	1988 (pf)	%age reduction (-) or increase (+) in the pf over pi
Control (untreated)	788.9	1251.4	1334.9	1107.4	+40.4
Phorate 10 G @ 1.5 g a.i./vine	815.8	927.6	531.1	296.7	-63.6
Phorate 10 G @ 3.0 g a.i./vine	903.9	(-25.9)	(60.2)	(-73.2)	
Bavistin 0.2% @ 5 l/vine	908.6	596.2	430.6	381.1	-57.8
Phorate 10 G @ 3 g a.i./+		(-52.3)	(-67.0)	(-65.6)	
Bavistin 0.2% @ 5 l/vine	728.9	1298.0	1451.8	1530.0	+68.4
Neemcake @ 2 kg/vine	760.7	(+3.7)	(+8.8)	(+38.2)	
C.D. at 5%	NS	627.1	449.4	380.2	-47.8
		(-49.9)	(-66.3)	(-65.7)	
		477.3	284.7	229.8	-69.8
		(-61.9)	(-78.7)	(-79.3)	
		576	570	625	

pi = Initial population, pf = Final population

Figures in parentheses denote percent reduction (-) or increase (+) over untreated control

M. incognita and *R. similis* in the roots estimated during different years are given in Tables I and II respectively. Final population (pf) of root knot nematode was significantly reduced over initial population (pi) in the vines treated with phorate 10 G at both dosages alone or in combination with bavistin soil drenching and neem cake ranging from 47.8 to 69.8 per cent (Table I). Maximum reduction in the nematode population (69.8%) was observed in the vines treated with neem cake followed by phorate 10G 3 g a.i./vine. On the contrary, the population of root knot nematode had significantly increased in the vines treated with bavistin alone(68.4%) followed by untreated control (40.4%). Similar trend was observed with regard to the population of *R. similis* (Table II). Phorate at both the dosages significantly reduced the nematode populations. Nematode population has considerably increased in the vines treated with bavistin alone which was more than the population increase in the untreated control. However, neem cake showed less efficacy in suppressing the population of *R. similis*.

Though there was significant reduction in the population levels of both the nematode species in the roots of pepper vines treated with phorate/neem

case, there was no corresponding decrease in the foliar yellowing indices (Table III) and subsequent improvement in the health of the vines. Further the disease manifested severely during the course of three years of experimentation leading to death of vines

ranging from 16.6 to 58.3 per cent in different treatments. The highest mortality of the vines was observed in the bavistin-treated vines alone with the highest percentage of positive isolation of *P. capsici* from the roots. This might be due to the fact that bavistin

Table II. Population of burrowing nematode, *Radopholus similis* in black pepper (per 5g of root)

Treatment	1985 (pi)	1986	1987	1988 (pf)	% reduction (-) or increase (+) in the pf over pi
Control (untreated)	462.2	797.3	889.6	682.7	+ 71.0
Phorate 10 G @ 1.5 g a.i./vine	351.2	270.8	143.7	162.8	- 45.0
		(-66.0)	(-88.8)	(-76.2)	
Phorate 10 G @ 3 g a.i./vine	413.3	236.1	49.6	94.5	- 69.7
		(-70.4)	(-94.4)	(-86.2)	
Bavistin 0.2% @ 5 l/vine	312.8	535.3	603.9	1107.6	+ 139.3
		(-32.9)	(-32.1)	(+62.2)	
Phorate 10 G @ 3 g a.i. + Bavistin 0.2% @ 5 l/vine	248.9	246.8	179.5	181.5	- 18.5
		(-69.1)	(-79.8)	(-73.4)	
Neemeake @ 2 kg/vine	343.7	388.7	204.4	396.5	- 4.1
		(-51.3)	(-77.0)	(-41.9)	
C.D. at 5%	N.S.	280	349	377	

pi = Initial population, pf = Final population

Figures in parentheses denote percent reduction (-) or increase (+) over untreated control

Table III. Foliar yellowing and mortality of black pepper in the experimental plot

Treatments	Foliar yellowing index				Mortality of the vines			% isolation of <i>P. capsici</i> (out of 50 root bits tested)
	1985	1986	1987	1988	No.	%		
Control (Untreated)	1.64	1.42	1.89	1.92	10	27.7	48	
Pherate 10G @ 1.5 g a.i./vine	1.67	1.39	1.28	1.66	6	16.6	40	
Phorate 10G @ 3 g a.i./vine	1.64	1.11	1.19	1.69	16	43.4	76	
Bavistin 0.2% @ 5 l/vine	1.61	1.18	1.81	2.24	21	53.3	60	
Phorate 10G @ 3 g a.i. + Bavistin 0.2% @ 5 l/vine	1.61	1.31	1.16	1.51	10	27.7	36	
Neemeake @ 2 kg/vine	1.53	1.22	1.45	1.42	7	19.4	48	

Foliar yellow on 0-3 scale (0=Healthy; 1=mild; 2=medium & 3=severe)

is known to suppress fungi like *Fusarium* sp. only and not *Phytophthora*. This also indirectly indicates that *Fusarium* sp. has no role in root rot and death of vines. The increased vine death in bavistin-treated vines might be due to its ineffectiveness to control *Phytophthora*. Further studies are needed to understand the microbiological changes in the soil in relation to *Phytophthora* population to see whether there is any disturbance in the native population of natural antagonists of *Phytophthora* present in bavistin-treated plots. All the vines irrespective of treatment gave positive isolation of *Phytophthora* indicating its role in causing root damage and subsequent expression of symptoms. Anandaraj *et al.* (1988) indicated that when the infection of *P. capsici* is restricted to certain portion of the root system, initially the vines express symptoms such as foliar yellowing and defoliation. These studies indicated that infection by plant parasitic nematodes and fungi could result in root degeneration leading to slow decline. Since spatial segregation of plant parasitic nematodes and *Phytophthora* cannot exist under field condition, the study strongly suggests for an integrated disease management to check plant parasitic nematodes and *P. capsici* and to boost up the health and vigour of black pepper vine.

ACKNOWLEDGEMENTS

The authors are thankful to Dr. M.K. Nair, Director, Central Plantation Crops Research Institute, Kasaragod and Joint Director, CPCRI Regional Station, Vittal for providing facilities and to Dr. K.B. Abdul Khader, Sr. Scientist (Agronomy), CPCRI Regional Station, Vittal, for all the help given throughout. Thanks are due to Dr. A. Ramadasan, Director, National Research Centre for Spices, Calicut for the encouragements.

REFERENCES

- ANANDARAJ, M., RAMACHANDRAN, N. and SARMA, Y.R. 1988. Epidemiology of foot rot disease of black pepper (*Piper nigrum* L.) in India. Paper presented at the International Workshop on Wilt Diseases of Black Pepper, 27-29 October, 1989, Panaji, Goa.
- ANONYMOUS. 1985. Central Plantation Crops Research Institute, 1985. Annual Report for 1984, Kasaragod, India, p. 16.
- BRIDGE, J. 1978. Plant nematodes associated with cloves and black pepper in Sumatra and Bangka, Indonesia. (ODM Technical Report on visit to Indonesia, 0-19 July, 1978). Ascol, Berks, U.K., Ministry of Overseas Development (1978). iii + 19 pp. (En) Imperial College of London University, Silwood Park, Sunninghill, Ascot, Berks, U.K.
- CHRISTIE, J.R. 1957. The yellow disease of pepper and spreading decline of citrus. *Plant Dis. Rept.* 41: 267-268.
- HUBERT, F.P. 1957. Diseases of some export crops in Indonesia. *Plant Dis. Rept.* 41 : 55-64.
- ICHINOHE, M. 1975. Infestation of black pepper vines by the root knot nematode, *Meloidogyne incognita* at Tom-Aeu, Para, Brazil. *Jap. J. Nematol.* 5 : 36-40.
- ICHINOHE, M. 1976. Nematode problems of black peppers in Bangka Island, Indonesia. *Nematology Newsletter* 22 : 2.
- MUSTIKA, I. 1978. An observation on the relationship between nematode population and yellow disease on black pepper in Barcka. Pemberitaan Lembaga Penelitian Tanaman-Industri, Bogor, Indonesia No. 30 11-22.
- NAMISAR, K.K.N. and SARMA, Y.R. 1977. Wilt diseases of black pepper. *J. Plantn. Crops.* 5 : 92-103.
- NAMISAR, K.K.N. and SARMA, Y.R. 1979. Factors associated with stem wilt of pepper. Proc. PLACROSYM-2, 1979, pp. 343-353.
- RAMANA, K.V., MCHANDRAS, C. and BALAKRISHNAN, R. 1987. Role of plant parasitic nematodes in the stem wilt disease complex of black pepper (*Piper nigrum* L.) in Kerala. *India J. Nematol.* 17 : 225-230.
- TING, W.P. 1975. Plant Pathology in Peninsular Malaysia. *Rev. Pl. Path.* 54 : 207-305.
- VECHT, J. Van Der. 1950. Op Planten parasiterende aaltjes (Tylenchidae). In *De rassen van de cultuurgewassen in Indië*, p. 16-42, 'S Gravenhage, W. van Hoeve.
- VENKITESAN, T.S. and SETTY, K.G.H. 1977. Pathogenicity of *Gudephelus similis* to black pepper (*Piper nigrum*). *Indian J. Nematol.* 7 : 17-26.