Ages, rosemary was associated with wedding ceremonies; either the bride would wear a rosemary head piece, the groom and guests would wear a sprig of rosemary or the couple would plant a rosemary bush on their wedding day. In Hungary, ornaments made of rosemary were once used as a symbol of love, intimacy and fidelity of a couple.

Some of these myths and beliefs had an underlying scientific logic behind them, as present-day studies reveal. Now it is clear that the essential oil and tannins present in rosemary produce an aromatic smoke with cleansing and purifying properties.

#### 25.1.1 Origin and distribution

Rosemary (*Rosmarinus officinalis* L.) syn. Compass plant, Family Lamiaceae, is native to the Mediterranean – from Spain to the Balkans and into North Africa (Giugnolinini, 1985). At present, it is widely cultivated in Spain, Morocco, Tunisia, France, Algeria, Portugal and China and, to a limited extent, in India in the Nilgiris and Bangalore. Rosemary chemotypes with distinct morphological characters and oil quality occur in Italy (Mulas *et al.*, 1998).

#### 25.1.2 Description

Rosemary is a dense, evergreen, hardy perennial aromatic herb of 60-200 cm in height with small (2–4 cm) pointed, leaves (Fig. 25.1). The upper surface of the leaf is dark green or blue, while the underside of the leaf is white; the leaves are resinous. Branches are rigid with fissured bark and stems brown, square and woody. Flowers are whitish, blue or purple in cymose inflorescence. The leaves, flowering tops and twigs yield an essential oil and oleoresin valued in recipes, traditional medicine, modern medicine and aromatherapy as well as in the perfume and flavour industries. Although it is generally *R. officinalis* that is used for oil extraction, in Morocco *R. eriocalyx* is also used for extracting essential oil (Elamrani *et al.*, 2000).

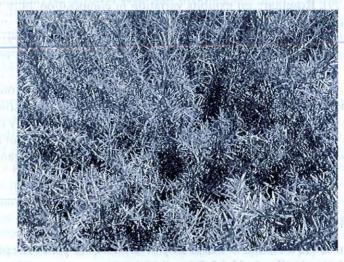


Fig. 25.1 Rosemary.

25

# Rosemary

B. Sasikumar, Indian Institute of Spices Research, India

**Abstract**: Rosemary is an aromatic herb that has been known from ancient times as a memory herb. This chapter briefly describes the myths and folklore that are associated with this plant, and goes on to give details of the agricultural techniques used in its production, including biotechnology and developments in post-harvest processing and analytical techniques that have improved the extraction of rosemary oil and oleoresin. The culinary and medical uses of the herb are also outlined in this chapter, as is the use of rosemary as a herbal pesticide. The toxicology of rosemary and its oils is discussed at the end of the chapter, which concludes with suggestions for key topics that require ongoing research.

Key words: biomedical uses, chemical profile, oil extraction, pesticide, post-harvest technology, rosemary.

### 25.1 Introduction

Rosemary, the memory herb of the mint family, has been used by man since ancient times. Records of the use of rosemary appear in cuneiform on Sumerian stone tablets of the fifth millennium BC. The Chinese and Greeks used rosemary as a health conditioner; the Greeks, who wore rosemary wreaths in their hair, also believed that rosemary strengthened the brain and enhanced memory. In Egypt, the herb was buried with the pharaohs. Hungary water, an infusion of rosemary in spirits of wine, is thought to have been first prepared for Queen Izabella of Hungary in 1235 to renovate her paralysed limbs. The word rosemary is derived from the Latin word '*rosmarinus*', meaning 'sea dew'. It was also called '*antos*' by the ancient Greeks, meaning the flower of excellence or 'libanotis' for its smell of incense (Giugnolinini, 1985).

There are many myths and much folklore associated with rosemary. It is believed that placing rosemary sprigs under the pillow would ward off evil spirits and nightmares from the sleeper and that the aroma of rosemary would keep old age at bay (Rose, 1974). During the Middle Ages it was believed that burning rosemary leaves and twigs would scare away evil spirits and disinfect the surroundings; later, it was burned by judges to protect against illness brought in by prisoners. It was also known as a symbol of fidelity and in Europe wedding parties burned rosemary as incense. Rosemary has also been used as a symbol of love and loyalty. As such, in the Middle

#### Table 25.1 Composition of dried rosemary leaves

Moisture	5.7 %
Protein	4.5 %
Fat	17.7 %
Crude fibre	19.0 %
Carbohydrates	47.4 %
Ash	6.0 %
Calcium	1.5 %
Phosphorous	0.7 %
Iron	0.03 %
Sodium	0.004 %
Potassium	1.0 %
Vitamin A	175 IU/100 g
Vitamin B	0.51 mg/100 g
Vitamin C	61.3 mg/100 g
Energy	440 calories/100 g

#### Source: Farooqi et al., (2005).

The annual global oil production is 200–300 metric tonnes. Many different varieties and cultivars are grown, each varying in flower colour (blue/pink/white), plant habit (erect/spreading), leaf colour (olive/blue/green), size of leaves, etc.

#### 25.1.3 Chemical composition

The composition of dried rosemary leaves is shown in Table 25.1. The main components of rosemary oil are 1,8-cineol (15-20%), camphor (15-25%), borneol (16-20%), bornyl acetate (up to 7%),  $\alpha$ -pinene (25%); in addition, the oil contains minor amounts of  $\beta$ -pinene, linalool, camphene, subinene, myrcene,  $\alpha$ -phellandrene,  $\alpha$ -terpinene, limonene, *p*-cymene, terpinolene, thujene, copalene, terpinen-4-ol,  $\alpha$ -terpineol, caryophyllene, methyl chavicol and thymol. The initial distillation fraction contains mostly  $\alpha$ -thujene,  $\alpha$ -pinene, camphene,  $\beta$ -pinene and 1,8-cineol, while camphor and bornyl acetate constitute the bulk of the later distillation (Prakasa Rao et al., 1999). Rosemary oil exhibits variation in composition, both profile and percentage of each component, depending on growing location and/or other factors such as source population, fertilizers and phenology (Boyle et al., 1991; Arnold et al., 1997; Boutekedjiret et al., 1999; Ouahada, 2000; Porte et al., 2000; Wolski et al., 2000; Guazzi et al., 2001). Pintore et al., (2002) reported a total of 58 compounds from rosemary oil from Sardinia and Corsica (Italy) based on gas chromatography retention index (GC-RI), GC mass spectroscopy (GC-MS) and carbon nuclear magnetic resonance (C-NMR) studies. A study of Moroccan rosemary oil not only demonstrated the existence of three different rosemary chemotypes but also identified a total of 91 compounds, based on GC and GC-MS studies (Elamrani et al., 2000).

## 25.2 Production and cultivation of rosemary

Rosemary can be grown either as a field crop or as an indoor plant. The plant thrives well in well-drained soils of pH 6.5–7.0 and with warm, sunny weather conditions

(Doulgas, 1971). It will also grow in a semi-arid tropical climate. The plant is, however, susceptible to severe cold and frost, although frost-resistant varieties are now available (Domokos et al., 1997). Male sterility is known to occur in natural populations of rosemary. Mitochondrial genome polymorphism linked to male sterility has been reported from Spain (Hidalgo-Fernandez et al., 1999). This male sterility may be responsible for spontaneous population evolution and the occurrence of chemotypes in rosemary. Rosemary is a perennial herb propagated either by cuttings or seeds. Cuttings of 10–15 cm length from selected mother plants are ideal for vegetative propagation of rosemary. Treatment of cuttings with growth hormones such as indole butyric acid (IBA), indoleacetic acid (IAA) or saponin are reported to enhance rooting of cuttings (Shah et al., 1996; Silva and Pedras, 1999). The end of winter has been found to be the most successful season for rooting cuttings (Silva and Pedras, 1999). Cuttings with the lower leaves removed are first planted in raised sand beds in a protected nursery. Regular watering is needed to achieve good sprouting. It then takes about 45-50 days for the cuttings to be transplanted to the main field.

Rosemary seeds are very small and black in colour. In India, the seed nursery is usually in operation from September to November. Seed rate is about 0.2–2.5 g seed 1 m<sup>2</sup> area (Farooqi and Sreeramulu, 2001). Raised seed beds with adequate shade, sufficient watering, good drainage and weeding ensure healthy seedlings. Seedlings are transplantable at 8–10 weeks; the usual spacing adopted for the rosemary plants is  $45 \times 45$  cm as a monocrop.

Soil properties are known to influence the yield and composition of the rosemary oil harvested. Moretti *et al.* (1998a) reported that granite silt soils result in a better herb yield and oil quality when compared with calcareous soils. A fertilizer dose per hectare of 40 kg  $P_2O_5$ , 40 kg  $K_2O$  and 20 kg N with 20 metric tonnes farmyard manure is recommended for rosemary production in India (Farooqi and Sreeramulu, 2001). Furthermore, N level can be raised to 300 kg ha<sup>-1</sup> in different splits to maximize oil yield. Close spacing coupled with increased nitrogen dose result in higher herbage and oil yield (Prakasa Rao *et al.*, 1999). Studies conducted in Italy (Sardinia) on fertilizer dose and weed management revealed that applying 80 kg N + 60 kg  $P_2O_5$  ha<sup>-1</sup> coupled with hand weeding of the major weeds such as *Genista corsica* and *Cytisus* spp. increased herbage yield and oil (Milia *et al.*, 1996). Biofertilizers, such as *Azospirillum*, *Azotobacter* and vesicular arbuscular mycorrhiza (VAM), applied in combination with inorganic fertilizers, have also been reported to have a beneficial effect on the herb and oil yield of rosemary (Anuradha *et al.*, 2002).

Weed control in rosemary is achieved through occasional hand weeding and intercultivation. Other husbandry techniques that have been studied include foliar spraying of iron, use of growth regulators, irrigation and pruning. The application of iron as a foliar spray in irrigated rosemary increased the verbenone concentration in the oil (Moretti *et al.*, 1998b). Studies on the effect of growth regulators (brassinosteroids and uniconazole) on growth, yield and chemical composition of rosemary in Egypt revealed the beneficial effect of growth hormones (Tarraf and Ibrahim, 1999). Irrigation is reported to be beneficial for herbage yield of rosemary. To establish the crop well in the field, two irrigations per week are recommended in India. Subsequently, irrigation once a week will be sufficient (Farooqi

© Woodhead Publishing Limited, 2012

and Sreeramulu, 2001). Drip fertigation with water-soluble fertilizers (80% WSF) coupled with micronutrients is reported to increase growth, yield and quality traits of rosemary (Vasundhara et al., 2002). Field experiments on water requirements of rosemary in Egypt revealed that irrigation once every 14 days resulted in high herbage and oil yield (Kandeel, 2001). A study on the effect of soil moisture regime, irrigation water (IW): cumulative pan evaporation (CPE) ratios and nitrogen level on herbage and oil vield of rosemary on alfisols in Bangalore. India, indicated that a soil moisture regime maintained at 0.50 IW:CPE ratio with 150 kg N ha<sup>-1</sup> significantly increased herbage and oil yield of rosemary (Singh and Ramesh, 2000). The correct soil moisture/nutrient regime is significant as unstressed rosemary plants are reported to yield higher levels of essential oil and phenolic compounds than those subjected to water/nutrient stress (Solinas et al., 1996). Pruning of rosemary is advisable, though not essential, after 2 or 3 years to enhance shoot and leaf production (Faroogi and Sreeramulu, 2001). As rosemary is a perennial herb, aged plantations (10-12 years old) need to be rejuvenated by cutting back the plant to a height of 4-5 cm above ground coupled with fertigation for better herbage yield.

#### 25.2.1 Diseases and pests

The important diseases affecting rosemary are: collar and root rot (Phytophthora incognitae, P. drechsleri), blight (Rhizoctonia spp., Phytocoris rosmarini and Orthotylus ribesi), foliar necrosis/leaf spot (Alternaria alternata), aerial blight (Ralstonia solani AG-4), hook disease (Botrytis cinerea) and powdery mildew (Oidium spp.) (Perello and Dal Bello, 1995; Minuto and Garibaldi, 1996; Cacciola et al., 1997; Conway et al., 1997; Villevieille et al., 1999). Alternaria leaf spot will be severe in humid and less well-ventilated areas, whereas powdery mildew can be severe in shaded conditions. Aerial blight is a major disease in greenhouse-grown rosemary. Various crop husbandry measures can reduce the severity of the impact of these diseases: (i) harvesting before blooming will help to restrict the crop loss due to hook disease; (ii) improved drainage can reduce the risk of root rot; (iii) spraying and drenching with the fungicide Maneb (1%) are effective against root rot and blight; (iv) sulphur dusting is recommended against powdery mildew. Biological control agents have also been investigated for use on rosemary. Biocontrol of powdery mildew with a commercial formulation of Ampetomyces quisqualis gave partial control (Minuto and Garibaldi, 1996). An isolate of the biocontrol agent, Laetisaria arvalis, if incorporated in the pots, followed by foliar spray of fungicides at a low dose, was found to check aerial blight of rosemary better than separate applications of fungicide or biocontrol agent alone (Conway et al., 1997). Biocontrol agents such as Trichoderma spp., and fluorescent pseudomonads are worth trying, as there is a premium price for organically produced herbs. These biocontrol agents can be components in an integrated disease management approach for rosemary. However, in the case of aerial blight of rosemary no synergistic effect of soil amendment with Trichoderma harzianum and foliar spray with iprodione (fungicide) was observed (Conway et al., 1997). Mites are a serious insect pest in rosemary grown in India (Farooqi et al., 2005). Spraying of Ethion (0.05%) is recommended to control the infestation.

#### 25.2.2 Biotechnology

*In vitro* studies of rosemary are in progress in several laboratories worldwide. Published research has focused on several topics: the callus induction potential of various explants from rosemary; the essential oil profile of *in vitro* cultures under salt stress; the *in vitro* production of the pigment shisonin; and the *in vitro* production of carnosic acid in callus cultures and regenerated shoots (Zhu-Ru *et al.*, 1996; Tawfik, 1997; Yang-Rong Hui *et al.*, 1997; Hashimoto *et al.*, 1997; Caruso *et al.*, 2000). However, despite the ongoing research, large-scale downstream processing of valueadded products of rosemary using these *in vitro* techniques, or scale-up of the process for commercial exploitation, is still in its infancy.

### 25.3 Post-harvest technology and further processing

Rosemary leaves, flowering tops, flowers and twigs are of economic importance. The first harvesting is carried out about eight months after planting, with the onset of flowering or just before flowering. In the first year, two crops can be taken, whereas in the subsequent years two to four harvests are possible at an interval of 100-120 days. Generally, harvesting of the plants can be carried out until 50% flowering. At above 90% flowering, harvesting is not desirable (Farooqi and Sreeramulu, 2001). Tender, non-hardy shoots are also harvested for distillation upon attaining full size. Usually the harvested leaves, flowering tops and shoots are used for downstream processing without drying. However, the leaves and twigs can also be used after drying for oil extraction. Drying studies in normal conditions at 50°C and in dehydrated air (30°C) revealed that in terms of percentage values of the characteristic volatile oil compounds, the dehydrated air dried product is on par with raw leaves (di Cesare et al., 2001). However, Ibanez et al., (1999) reported that, of the different drying methods, the traditional method (drying in a ventilated room) was the best in terms of the yield and quality of the antioxidant principles of rosemary. The genotype of the plant, the age of the leaf and growing conditions are known to affect the oil quality, especially the quantities of the antioxidant principle, carnosic acid, in the oil (Hidalgo et al., 1998). Boutekedjiret et al., (1999) reported a phenology-dependent variation in yield and quality. For best oil yield, the flowering stage of rosemary is preferable, although the oil quality from plants at this growth stage is slightly inferior. To achieve high yields of good-quality oil, the grower has to select an appropriate growth stage when harvesting.

## 25.3.1 Extraction of rosemary oil

Rosemary oil is colourless to pale yellow and its properties are shown in Table 25.2. Rosemary oil is usually recovered by steam or water distillation, although supercritical fluid extraction (SFE) using CO<sub>2</sub> as a solvent is also now being used (Coelho *et al.*, 1997; Bicchi *et al.*, 2000). Detailed studies aimed at standardizing the various parameters for supercritical CO<sub>2</sub> extraction of the oil – such as pressure, temperature, particle size, CO<sub>2</sub> rate, etc. – are in progress (Carvalho *et al.*, 2005; Bensebia *et al.*, 2009; Ivanovic *et al.*, 2009; Zermane *et al.*, 2010). A new extraction process, the Fast Control Pressure Drop method, has also been reported (Rezzoug *et al.*, 2005).

 Table 25.2
 Physical properties of rosemary oil

Solubility (90 % ethanol)	1 vol. (EOA)	
Total alcohol as borneol	Minimum 8 %	
Esters as bornyl acetate	Minimum 1–5%	
Refractive index (20°C)	1.464-1.476	
Optical rotation (20°C)	$-5^{\circ}$ to $+10C$	
Specific gravity (25°/25 C)	0.894-0.912	

This process involves exposing the rosemary leaves for a brief period of time to steam pressure varying from 0.5-3 bar followed by an instantaneous decompression to vacuum (about 15 mbar). Most of the oil (90%) comes out within the first 60 minutes of the conventional distillation, although distillation can be continued for 120 minutes for full recovery of rosemary oil under field distillation conditions (Prakasa Rao *et al.*, 1999).

It has been found that the oil obtained from leaves and flowering tops is of better quality than the oil obtained from whole-plant distillation. It has also been reported that, in terms of the yield and quality profile of the rosemary oil, steam distillation is preferable to water distillation (Boutekedjiret *et al.*, 1997). Blanching (microwave blanching for 1 min) is observed to have a positive effect on retention of the antioxidant principles, green colour and texture of rosemary; however, blanching leads to total loss of volatile oils (Singh *et al.*, 1996).

The oils and extracts obtained from rosemary by steam distillation, water distillation and controlled instantaneous decompression were analysed by GC and GC– MS in order to determine their chemical composition (Boutekedjiret *et al.*, 2004). It was observed that steam distillation seemed to be the most appropriate process for oil isolation of rosemary since it gave the highest yield, and an oil composition similar to values quoted in the literature and in commercial standards. The composition and antioxidant properties, as well as antimicrobial activities, of oil extracted using supercritical CO<sub>2</sub> have also been confirmed (Genena *et al.*, 2008).

The oil content of fresh rosemary leaves is 1% and in shade-dried leaves it increases to 3% (Farooqi and Sreeramulu, 2001). In theory, the yield from 1 ha is approximately 10–12 tonnes of herbs per year, yielding 25–100 kg oil (Farooqi and Sreeramulu, 2001). However, in field distillation conditions, the oil yield varies from 0.5–0.9% (Prakasa Rao *et al.*, 1999), so annual yields may be lower.

#### 25.3.2 Extraction of active compounds

For the extraction of the active compounds of rosemary, conventional solvent extraction techniques using solvents such as hexane, benzene, ethylene, chloroform, dioxane and methanol (Chang *et al.*, 1977) besides distillation and SFE are routinely employed. SFE using  $CO_2$  has been reported to be superior to liquid solvent sonication for the maximum recovery of carnosic acid in its pure form (Tena *et al.*, 1997). Recent research on improving the yield and quality of rosemary extract with new techniques has shown encouraging results. Superheated water under pressure between 125 and 175 °C has been shown not only to extract high-quality oxygenated fragrance and flavour compounds from rosemary rapidly but also to produce higher

yields than steam distillation (Basile *et al.*, 1998). Ibanez *et al.*, (1999) proposed a two-step SFE and fractionation of essential oil-rich oleoresin and antioxidant compounds by varying the pressure and temperature requirements. Enzyme-assisted ensiling (ENLAC) prior to polyphenol extraction is reported to double the yield of polyphenol from rosemary (Weinberg *et al.*, 1999).

Rosemary oleoresin, also known as rosemary oil extract, is a green to brownish green semi-olid to pasty extract which is fluid when warmed to 50 °C. Different grades of rosemary oleoresin are available, with the grade being dependent on the volatile oil concentration, in particular the carnosic acid concentration (Heath, 1981). It is the carnosinic acid that is believed to be responsible for the potent anti-oxidant properties of rosemary oleoresin. These properties will be discussed in a later section of the chapter.

#### 25.4 Main uses of rosemary

#### 25.4.1 Culinary uses and food processing

Rosemary leaves and flowering tops have many culinary uses: lamb roast, mutton preparations, marinades, bouquet garni, baked fish, rice, soups, salads, occasionally with egg preparations, dumplings, apples, summer wine cups and fruit cordials, in vinegar and oil (Bonar, 1994). The fresh and dried leaves of rosemary are used frequently in traditional Mediterranean cuisine as they have a bitter astringent taste and are aromatic. Dried and powdered leaves are added to cooked meat, fish, poultry soup, stews, sauces, dressings, preserves and jams. Mixed with sage, the leaves are also used in pork preparations. When burned the leaves give off a distinct mustard smell which is favoured in barbecuing.

Rosemary is a most effective herb with a wide range of uses in food processing. In Europe and the USA, rosemary is commercially available for use as an antioxidant, though not technically listed as natural preservative or antioxidant, especially in Europe (Yanishlieva-Maslarova and Heinonen, 2001). Rosemary has potential applications in the suppression of warmed over flavour (WOF) (Valenzuela and Nieto, 1996). The main antioxidant principles in rosemary are carnosic acid, 12-methoxy carnosic acid and carnosol, as well as the antioxidative diterpenes such as epirosmarinol, isorosmanol, rosmaridiphenol, rosmariquinone and rosmarinic acid (Richheimer et al., 1996). The antioxidant properties of rosemary are attributed to its ability to scavenge superoxide radicals, lipid antioxidation, metal chelating, etc. Extracts and essential oil of rosemary can be used to stabilize fats, oils and fatcontaining foods, e.g. butter, against oxidation and rancidity (Zegarska et al., 1996; Pokorny et al., 1998) and also to stabilize fermented meat products (Korimova et al., 1998). A useful review by Yanishlieva-Maslarova and Heinonen (2001) discusses the literature on rosemary and sage antioxidant principles, covering their extraction, properties and applications, and the chemistry involved. Deodorized liquid, commercial antioxidant formulations of rosemary, either as mono-herbal or as polyherbal (rosemary, thyme, sage, oreganum) formulations, i.e. 'Herbor 025', 'Spice Cocktail', etc., are available (Aruoma et al., 1996).

Rosemary oil also has applications in meat preservation. The addition of rosemary oleoresin to ground chicken had an overall positive effect on raw meat

© Woodhead Publishing Limited, 2012

appearance during storage and on the flavour of the cooked meat (Koekamnerd *et al.*, 2008). The antioxidative properties of rosemary oleoresin and its inhibition of off-flavours in pre-cooked roast beef slices have also been reported (Murphy *et al.*, 1999).

#### 25.4.2 Aromatherapy and cosmetics

Rosemary oil is used in aromatherapy to stimulate the central nervous system and blood circulation, to relieve muscular pain. Add a few drops of rosemary essential oil to any massage oil, such as sweet almond oil, blend together and massage gently into the area affected by pain.

It also has beneficial effects in promoting hair growth. Massaging a cocktail of essential oils of rosemary, thyme, cedar wood and lavender in carrier oils of jojoba and grape seed over a period of 7 months into the scalp of patients with *Alopecia areata* was found to be a safe and effective treatment (Hay *et al.*, 1998). Rosemary oil stimulates hair follicles and circulation in the scalp and thus may also be helpful in premature baldness (http://www.holistic-online.com/Herbal-Med/\_Herbs/ h228.htm). Rosemary is used in curing acne (Baumann, 2007).

In addition to its use in aromatherapy, rosemary also has several cosmetic applications. When used in a shampoo, rosemary oil can help to control dandruff and greasy hair; it is also known to impart a black colour to hair. Rosemary cleansing cream can be used for oily skin (Bonar, 1994). Rosemary oil is a component in soaps, room fresheners, deodorants, perfumes, skin lotions, etc., either in formulations with other herb oils or singly. The traditional Hungary water is a perfume based on rosemary oils. The flower, calyx and leaves of rosemary are used in potpourri and herb pillows (Bonar, 1994).

#### 25.4.3 Traditional medicinal uses

Hungary water prepared from rosemary is considered to be a revitalizing agent and the herb has an old reputation for improving memory. The presence of rosemary in one's body is believed to enhance clarity of mind and memory, akin to the belief surrounding sweet flag (Acorus calamus) in India. In traditional medicine, herbalists recommend rosemary oil for a wide range of disorders including pulmonary diseases. Rosemary oil has been described as having the following properties: stomachic, wound healing (poultice), choleretic, antidiabetic, diuretic, antidepressant and antispasmodic (Oury, 1984; Giugnolinini, 1985; Erenmemisoglu et al., 1997). The whole plant, in the form of decoction, infusion, extract in ethanol (for external application) and essential oil, is administered against digestive disorders, vaginitis, leucorrhoea, respiratory diseases, varicose vein, heart pain, inflammation and dizziness by the native people of Mexico and Central America (Santos García-Alvarado et al., 2001). In Russia and the Central Asian countries of the former Soviet Union, leaves of rosemary preparation (gallenical and powder made into cigarettes) are used to treat asthma (Mamedov and Craker, 2001). The abortifacient (anti-implantation) effect of rosemary extract is also known (Lemonica et al., 1996). A distilled water obtained from rosemary flower is used as an eye lotion (Farooqi *et al.*, 2005).

## 25.4.4 Modern research into medicinal properties

Rosemary is today credited with having many varied medicinal properties. Modern analysis techniques have enabled the individual components of the oil to be identified and their effects investigated. The medicinal properties and the chemical constituents/part of the plant that are now thought to be involved in these effects are listed below:

- carminative properties flavanoids;
- antidepressant, antispasmodic properties volatile oils;
- rubefacient properties phenolics;
- antimicrobial properties diterpenes;
- emmenagogue properties oleanolic acid;
- anti-inflammatory properties carnosol;
- carcinogen blocking and liver detoxifying properties carnosol and whole-plant extract;
- antirheumatic properties ointment of rosemary oil;
- abortifacient properties (aqueous extract).

### HIV treatment

Liquid deodorized extract of rosemary ('Herbor 025') and oily extract of mixture of herbs such as rosemary, thyme, sage and oreganum ('Spice Cocktail') inhibited human immunodeficiency virus (HIV) infection at very low concentrations though they were cytotoxic. Carnosol and carnosic acid were found top be the main active constituents of the extracts (Aruoma *et al.*, 1996). Purified carnosol @ 8  $\mu$ M exhibited anti-HIV activity besides beingon-cytotoxic.

## Cardiovascular effects

The cardiovascular effects of rosemary extract on the isolated intact rabbit heart demonstrated significant positive inotropic effect and coronary vasodilatation (Khatib *et al.*, 1998). Administering an infusion of dried rosemary leaves resulted in decrease of blood glucose levels in normoglycaemic and diabetic mice and had no toxic effects (Erenmemisoglu *et al.*, 1997).

#### Cancer prevention

Rosemary is now gaining importance in cancer treatment. Rosemary has been shown to have potential as a source of anticancer molecules and as a treatment to enhance the bioavailability of cancer drugs (Plouzek *et al.*, 1999; Huang *et al.*, 2009; Rasha and Abdella, 2010; Yesïl-Celïktas *et al.*, 2010). The major anticancer compounds identified from rosemary are: carnosol, carnosic acid, ursolic acid, befulinic acid, rosmaridiphenol and rosemanol. Carnosol is found to reduce cellular nitric oxide in mice, a free radical that can damage DNA (Chan *et al.*, 1995). It is also reported, in rats, that administering a whole-plant extract of rosemary is more effective in preventing the carcinogen 7,12-dimethyl benz(a)anthracene (DMBA) from binding to breast cell DNA than administering carnosol or ursolic acid alone (Singletary *et al.*, 1996).

Extract of rosemary is found to increase the intracellular accumulation of the common chemotherapy drugs such as dexorubicin (DOX) and vinblastine (VIN) in

drug-resistant MCF-7 human breast cancer cells leading to increased availability of the drugs for bioactivity (Plouzek et al., 1999). Adding rosemary extract to cooked beef reduces the formation of heterocyclic amines, a carcinogen (Puangsombat and Smith, 2010).

#### Diuretic and other effects

Haloui et al., (2000) reported a diuretic effect of aqueous extract of rosemary (8% concentration) on Wister rats. Carnosic acid in rosemary is reported to shield the brain from free radicals, lowering the risk of strokes and neurodegenerative diseases like Alzheimer's and Lou Gehrig's (Lipton, 2007). Hyperglycaemic and insulin release inhibitory effects of the herb have also been reported (Al-Hader et al., 1994). Rosemary extract also prevents binding of aflatoxins to human liver cells and benzo(a)pyrenene to bronchial tissue (Offord et al., 1995).

#### Administration

Rosemary can be administered in the form of infusion, decoction, ethanol extract (external use), tinctures, rosemary wine, drug-containing volatile oil, powdered drug, liquid extract, dry extract and rosemary oil (mainly for external use). The infusion is prepared by adding 2 g of herb to 1 l of boiling water (Santos Garcia-Alvarado et al., 2001).

#### 25.4.5 Herbal pesticide

In the agricultural industry, rosemary is known to possess insect-repellent and antimicrobial properties which are beneficial in growing and storage of crops. Comparative laboratory studies have been carried out where different herbal powders. including rosemary powder, were dusted on stored grains of wheat and French bean; analysis of the results of these treatments on the pest species Sitophilus granarius and Acanthoscelides obtectus revealed that grain wheat can be very effectively protected against S. granarius by dusting the grain with rosemary powder (Kalinovic et al., 1997). Aphids are also repelled by the odour of rosemary and the mint Mentha pulegium. Based on GC-MS analysis, a few components of rosemary oil - i.e. 1.8cineole, D-1-camphor,  $\alpha$ ,1-camphor and  $\alpha$ -pinene – have been found to be the major principles repelling the aphids (Hori and Komatsu, 1997). The antirepellent property of rosemary oil to Myzus persiacea is attributed to 13 compounds: the most important being linalool, D-1-camphor and α-terpineol (Hori, 1998).

Rosemary oil (1%) is found to reduce the fecundity rate of the predacious mites Amplyseius zaheri and A. barkeri (Momen and Amer, 1999). The repellent and oviposition-deterring activities of rosemary oil on the spider mites Tetranychus usticae and Eutetranychus orientalis have also been reported (Amer et al., 2001). Essential oil of rosemary in vapour form is shown to reduce fecundity, decrease egg hatchability, increase neonatal larval mortality and adversely affect offspring emergence of Acanthoscelides obtectus (Papachristos and Stomopoulos, 2002).

Essential oil of rosemary has also been reported to have antimicrobial activity against an array of bacterial and fungal species including Listeria monocytogenes and Aspergillus niger (Baratta et al., 1998; Faliero et al., 1999). Gram-negative bacteria such as Staphylococcus aureus and S. epidermidis have been found to be more susceptible to rosemary oil than are other gram-negative bacteria such as Escherichia coli and Pseudomonas aeruginosa (Pintore et al., 2002). Rosemary leaves themselves are a source of antibacterial molecules; in laboratory studies, an active compound effective against the plant pathogen Streptomyces scabies has been isolated from the leaves of rosemary (Takenaka et al., 1997).

## 25.5 Toxicology and quality control

Rosemary is generally considered safe and devoid of toxic side-effects if taken in recommended doses. However, certain guidelines for its use should be followed. (i) rosemary oil if internally consumed or ingested in excess can trigger convulsions; (ii) pregnant and lactating women are advised not to use rosemary, as are people with epilepsy; (iii) rosemary oil should be used with caution by persons suffering from hypertension, blood pressure or insomnia; (iv) rosemary leaves in excess quantity can cause coma, spasm, vomiting and, in some cases, pulmonary oedema.

There have been occasional reports of allergic reactions such as skin irritation when rosemary has been applied externally. In addition, rosemary essential oil may have epileptogenic properties as a handful of case reports over the past century have linked its use with seizures in otherwise healthy children (Burkhard et al. 1999). Rosemary may also interfere with iron absorption; hence it should be avoided by persons with iron deficiency anaemia (Samman et al., 2001).

#### 25.5.1 Adulteration

Rosemary oil can be adulterated with white camphor oil, fractions of eucalyptus oil, Spanish sage oil, oil of turpentine and petroleum. The quality of rosemary oil can be ascertained based on its solubility in alcohol and by using an iodine test. Rosemary oil does not detonate with iodine but just dissolves with heating (Diete and Brannt, 1892).

### 25.6 Conclusion

Rosemary is emerging as an important herb, being a potential source of anticancer molecules, a functional food, a botanical nutraceutical and a functional pesticide. Despite the multifaceted importance of the herb, it has yet to receive adequate research attention. In many places, it is grown either as a minor herb on marginal lands or still grows wild. Varietal improvement, quality profiling including adulteration detection, toxicology and organic cultivation practices are some areas that require immediate attention, in addition to post-harvest technology.

## 25.7 References

AL-HADER A A, HASAN Z A and AQEL M B (1994) Hyperglycemic and insulin release inhibitory effects of Rosmarinus officinalis, J. Ethnopharmacol., 43(3): 217-21.

- AMER S A A, REFAAT A M and MAMEN F M (2001) Repellent and oviposition deterring activity of rosemary and sweet marjoram on the spider mites *Tetranychus urticae* and *Eutetranychus orientalis* (Acari. Tetranychidae), *Acta Phytopathol. Entomol. Hungaria*, **36**(1-2): 155-64.
- ANURADHA M N, FAROOQI A A, VASUNDHARA M, KATHIRESAN C and SRINIVASAPPA K N (2002) Effect of biofertilizers on the growth, yield and essential oil content in rosemary (*Rosmarinus* officinalis L.), Proc. National Seminar, Strategies for Production and Export of Spices, 24–26 Oct., Indian Institute of Spices Research, Calicut, Kerala.
- ARNOLD N, VALENTINI G, BELLOMARIA B and HOCINE L (1997) Comparative study of the essential oils from *Rosmarinus eriocalyx* Jordan & Fourr from Algeria and *R. officinalis* L. from other countries, *J. Essent. Oil Res.*, 9(2): 167–75.
- ARUOMA O I, SPENCER J P E, ROSSI R., AESCHBACH R., KHAN A., MAHMOOD W, MUNOZ A., MURICA A, BUTLER J and HALLIWELL B (1996) An evaluation of the antioxidant and antiviral action of extracts of rosemary and Provencal herbs, *Food Chem. Toxicol.*, **34**(5): 449–56.
- BARATTA M T, DORMAN H J D, DEANS S G, BLONDI D M and RUBERTO G (1998) Chemical composition, antimicrobial and antioxidative activity of laurel, sage, rosemary, oregano and coriander essential oils, J. Essent. Oil Res., 20(6): 618–27.
- BASILE A, JIMENEZ CARMONA M M and CLIFFORD A A (1998) Extraction of rosemary by superheated water, J. Agric. Food. Chem., 46(12): 5205–9.
- BAUMANN LS (2007) Less-known botanical cosmeceuticals, Dermatol. Ther., 20(5): 3303-42.
- BENSEBIA O, BARTH D, BENSEBIA B and DAHMAN A (2009) Supercritical CO<sub>2</sub> extraction of rosemary: effect of extraction parameters and modelling, J. Supercrit. Fluids, 49: 161–8.
- BICCHI C, BINELLO A and RUBIOLO P (2000) Determination of phenolic diterpene antioxidants in rosemary (*R. officinalis* L.) with different methods of extraction and analysis, *Phytochem. Anal.*, **11**(4): 236–42.
- BONAR A (1994) Herbs A Complete Guide to their Cultivation and Use. Tiger Books International, London.
- BOUTEKEDJIRET C, BENHABLES N E H, BELABBES R and BESSIERE J-M (1997) Effect of mode of extraction on yield and composition of the essential oil of *Rosmarinus officinalis*, *Rivista Italiana EPPOS*, **22**: 33–35.
- BOUTEKEDJIRET C, BELABBES R, BENTAHAR F and BESSIER J-M (1999) Study of *R. officinalis* L. essential oil yield and composition as a function of the plant life cycle, *J. Essent. Oil Res.*, **11**(2): 238–40.
- BOUTEKEDJIRET C, BELABBES R, BENTAHAR F, BESSIERE J-M and REZZOUG S A (2004) Isolation of rosemary essential oil by different process, J. Essent. Oil Res., 16(3): 195–9.
- BOYLE T H, CRAKER L E and SIMON J E (1991), Growing medium and fertilization regime influence growth and essential oil content of rosemary, *Hort. Sci.*, **26**(1): 33–4.
- BURKHARD P R, BURKHARDT K, HAENGGELI C A and LANDIS T (1999) Plant-induced seizures: reappearance of an old problem, J. Neurol., 246(8): 667–70.

CACCIOLA S O, PANE A, POLIZZI G and WATERHOUSE G M (1997) Collar and root rot of rosemary caused by two species of *Phytophthora*, *Informat. Fitopat.*, **47**(11): 35–42.

- CARUSO J L, CALLAHAN J, DE CHANT C, JAYASIMHULU K and WINGET G D (2000) Carnosic acid in green callus and regenerated shoots of *Rosmarinus officinalis* L, *Plant Cell Rep.*, 19(5): 500–503.
- CARVALHO JR R N, MOURA L S, ROSA P T V and MEIRELES, A A (2005) Supercritical fluid extraction from rosemary (*Rosmarinus officinalis*): Kinetic data, extract's global yield, composition and antioxidant activity, *J. Supercrit. Fluids*, **35**: 197–204.
- DI CESARE L F, VISCARDI D, FUSARI E L and NANI R C (2001) Volatile composition of fresh and dried rosemary, *Industrie Alimentari*, 40: 1343–5.
- CHAN M M-Y, HO C T and HUANG H I (1995) Effects of three dietary phytochemicals from tea, rosemary and turmeric on inflammation induced nitrite production, *Cancer Lett.*, 96: 23–9. CHANG S S, OSTIC-MATIJASAVIC B, HSIEH O A L and HUANG C L (1977) Natural antioxidants from rosemary and sage, *Food Sci.*, 42: 1102–6.

COELHO L A F, OLIVEIRA J V and PINTO J C (1997) Modelling and simulation of supercritical fluid extraction of rosemary essential oil, *Cie. Tec. Aliment.*, **17**: 446–8.

- CONWAY K E, MANESS N E and MOTES J E (1997) Integration of biological and chemical controls for *Rhizoctonia* aerial blight and root rot of rosemary, *Plant Dis.*, **81**(7): 795–8.
- DOMOKOS J, HETHELYI E, PALINKAS J, SZIRMAI S and TULOK M H (1997) Essential oil of rosemary (Rosmarinus officinalis L.) of Hungarian origin, J. Essent. Oil Res., 9(1): 41–5.
- DIETE C and BRANNT WT (1892) A Practical Treatise on the Manufacture of Perfumery. Applewood Books, Bedford, MA.
- DOUGLAS S J (1971) Cultivating condiments and seasonal plants for flavours and essential oil, *Flav. Ind.*, 222–5.
- ELAMRANI A, ZRIRA S, BENJILALI B and BERRADA M (2000) A study of Moroccan rosemary oils, J. Essent. Oil Res., 12(4): 487–5.
- ERENMEMISOGLU A, SARAYAMEN R and USTUN H (1997) Effect of *Rosmarinus officinalis* leaf extract on plasma glucose level in normoglycaemic and diabetic mice, *Pharmazie*, **52**(8): 645–6.
- FALERIO L, MIGUEL G M, GUERRERO C A C and BRITTO J M C (1999) Antimicrobial activity of essential oils of rosemary (*R. officinalis* L) and thyme (*Thymus mastichina* L), *Acta Hort.* (*ISHS*), **501**: 45–8.
- FAROOQI A A and SREERAMULU B S (2001) Cultivation of Medicinal and Aromatic Plants. Universities Press India Ltd, Hydrabad, 433–43.
- FAROOQI A A, SREERAMULU B S and SRINIVASAPPA K N (2005) Cultivation of Spice Crops. Universities Press India Ltd, Hyderabad.
- GENENA A K, HENSE H, SAMNIA JR A and SOUZA S M DE (2008) Rosemary (R. officinalis) a study of the antioxidant and antimicrobial activities of extracts obtained with super critical
- carbon dioxide, *Clenc. Tecnol. Aliment., Campinas*, **28**: 463–9. GIUGNOLININI L (1985) *Erbe Secondo Natura*. Secondo Natura Laboratorio Grafico, Vignate,
- Milano. GUAZZI E, MACCIONI S, MONTI G, FLAMINI G, CIONI P L and MORELLI I (2001) Rosmarinus officinalis
- in the gravine of Palagianello, Taranto, South Italy, J. Essent. Oil Res., 13(4): 231–3.
- HALOUI M, LOUEDEC L, MICHEL J B and LYOUSSI B (2000) Experimental diuretic effects of Rosemarinus officinalis and Centaurium erythraea, J. Ethnopharmacol., 71(3): 465-72.
- HASHIMOTO O, MIZUTANI H and NAKASHIMA R (1997) Production of shisonin in rosemary (Rosmarinus officinalis L.) tissue culture, Nippon Nogeikagaku Kaishi, 71(6): 605–56.
- HAY I C, JAMIESON M and ORMEROD A D (1998) Randomized trial of aromatherapy: successful treatment of Alopecia areata, Arch. Dermat., 134(11): 1349-52.
- HEATH H B (1981) Source Book of Flavours. AVI Book, Van Nostrand, New York. HIDALGO P J, UBERA J L, TENA M T and VALCARCEL M (1998) Determination of the carnosic acid content in wild and cultivated R. officinalis, J. Agric. Food Chem., 46(7): 2624–27.
- HIDALGO-FERNANDEZ P J, VICENTE P R, MALDONALDO J M and UBERA J J L (1999) Mitochondrial DNA polymorphism and gynodioecy in a natural population of *Rosmarinus officinalis* L., *Israel J. Pl. Sci.*, **47**(2): 77–83.
- HORI M (1998) Repellency of rosemary oil against *Myzus persicae* in a laboratory and in a screen house, *J. Chem. Ecol.*, **24**(9): 1425–32.
- HORI M and KOMATZU H (1997) Repellence of rosemary oil and its components against the onion aphid, *Neotoxoptera fermosana* (Takahashi) (Homoptera, Aphididae), *Appl. Ent. Zool.*, **32**(2): 303–10.
- HUANG H-C, HUANG C-Y, LIN-SHIAU S-Y and LIN J-K (2009) Ursolic acid inhibits IL-1 $\beta$  or TNF- $\alpha$ -induced C6 glioma invasion through suppressing the association ZIP/p62 with PKC- $\zeta$  and downregulating the MMP-9 expression, *Mol. Carcinogen.*, 48(6): 517-31.
- IBANEZ E, OCA A, DE MURGA G, LOPEZ SEBASTIAN S, TABERA J and REGLERO G (1999) Supercritical fluid extraction and fractionation of different pre-processed rosemary plants, *J. Agric. Food Chem.*, **47**(4): 1400–404.

- IVANOVIC J, DILAS S, JARDNIN M, VAJS V, BABOVIC N, PETROVIV S and ZIZONIC I (2009) Supercritical carbon dioxide extraction of antioxidant from rosemary (*R. officinalis* L.) and sage (*Salvia officinalis* L.), *J. Serb. Chem. Soc.*, **74**: 717–32.
- KALINOVIC I, MARTINCIC J, ROZMAN V and GUBERAC V (1997) Insecticidal activity of substances of plant origin against stored product insects, *Ochrana Rostlin*, **33**(7): 135–42.
- KANDEEL A M (2001) Effect of irrigation intervals on the growth and active ingredients of *Rosmarinus officinalis* L. plants, *Arab Univ. J. Agric. Sci.*, 9(2): 825–38.
- кнатів s, Alkofahi A, HASAN M and NAJIB N (1998) The cardio vascular effects of *Rosmarinus* officinalis extract on the isolated intact rabbit heart, *Fitoterapia*, **69**(6): 502–6.
- KOEKAMNERD T, ACON J C, HAN I Y and DAWSON P L (2008) Effect of commercial rosemary oleoresin in preparations on ground chicken thigh meat quality packaged in high oxygen atmosphere, *Poultry Sci.*, 87: 170–9.
- KORIMOVA L, MATE D and TUREK P (1998) The evaluation of raw fermented meat products stabilized with vitamin E and rosemary, *Folio Veterinaria*, **42**(4): 178–81.
- LEMONICA I P, DAMASCENO D C and SATSIDI L C (1996) Study of embryo toxic effects of an extract of rosemary (*Rosmarinus officinalis*), *Braz. J. Med. Biol. Res.*, **29**(2): 223–7.
- LIPTON SA (2007) Pathologically activated therapeutics for neuroprotection, *Nat. Rev. Neurosci.*, **8**(10): 803–8.
- MAMEDOV N and CRAKER L E (2001) Medicinal plants used for the treatment of bronchial asthma in Russia and Central Asia, J. Herbs, Spices Med. Plants, 8(213): 119–60.
- MILIA M, PINNA M E, SATTA M and SCARPA G M (1996) Preliminary examinations of agronomic aspects in Rosmarinus officinalis L. Seminario Internazionala di Chiusura del Progetto PIM, Sardagna, Cagliari, 18–19 Nov., 1994; Rivista Italiana EPPOS, 19: 125–30.
- MINUTO G and GARIBALDI A (1996) Comparison of methods of control of powdery mildew (*Oidium* spp.) on rosemary, *Informat. Fitopatol.*, **46**(12): 33–7.
- MOMEN M E and AMER S A A (1999) Effect of rosemary and sweet marjoram on three predacious mites of the family Phytoseiidae (Acari: Phytoseiidae), Acta Phytopathog. Entomolog. Hungaria, 34(4): 355–61.
- MORETTI M D L, PEANA A T, PASSINO G S and SOLINAS V (1998a) Effect of soil properties on yield and composition of *Rosmarinus officinalis* L. essential oil, J. Essent. Oil Res., 10(3): 261–7.
- MORETTI M D L, PEANA A T, PASSINO G S, BAZZONI A and SOLINAS V (1998b) Effect of iron on yield and composition of *Rosmarinus officinalis* L. essential oil, *J. Essent. Oil Res.*, **10**(1): 43–9. MULAS M, BRIGAGLIA N, CANI M R and SCANNERINI S (1998) Clone selection from spontaneous
- germ plasm to improve Rosmarinus officinalis L. crops, Acta Hort. (ISHS), 457: 287-94.
- MURPHY A, KERRY J P, BUCKLEY J and GRAY I (1999) The antioxidative properties of rosemary oleoresin and inhibition of off-flavours in precooked roast beef slices, J. Sci. Food Agric., **77**: 235–43.
- OFFORD E A, MACE K, RUFFIEUX C and PFEIFER A M (1995) Rosemary components inhibit benzo(a) pyrene induced geno toxicity in human bronchial cells, *Carcinogenesis*, **16**: 2057–62. OUAHADA s (2000) Tunisian rosemary oil, *Perf. Flav.*, **25**(6): 24–5.
- OURY P. (1984) Eucyclopedia des Plants et Fleurs Medicinale. Vol. IV. Editions Alliance, Paris. PAPACHRISTOS D P and STOMOPOULOS D C (2002) Repellent, toxic and reproduction inhibitory effect of essential oil vapour on Acanthoscelides obtecus (Say) (Coleptera: Bruchidae), J. Stored Prod. Res., 38(2): 117-20.
- PERELLO A and DAL BELLO G M (1995) Foliar necrosis caused by Alternaria on rosemary and Colletotrichum spp. on lavender, sage and marjoram, Invest. Agrar. Prod. Prot. Veg., 10(2): 275-81.
- PINTORE G, USAI M, BRADESI P, JULIANO C, BOATTOTOMI F, CHESSA M, CERRI R and CASANOV A J (2002) Chemical composition and antimicrobial activity of *R. officinalis* L. oils from Sardinia and Corsica, *Flavour Fragr. J.*, **17**(1): 15–19.
- PLOUZEK C A, CIOLINO H P, CLARKE R and YEH G C (1999) Inhibition of *p* glycoprotein activity and reversal of multi drug resistance *in vitro* by rosemary extract, *Eur. J. Cancer*, **35**(10): 154–5.

- PORTE A, GODOY R L, DE LOPES D, KOKETSU M, GONCLAVES S L and TORQUILHO H S (2000) Essential oil of *Rosmarinus officinalis* (rosemary) from Rio de Janeiro, Brazil, *J. Essent. Oil Res.*, **12**(5): 577–80.
- PRAKASA RAO E V S, GOPINATH C T, GANESHA RAO R S and RAMESH S (1999) Agronomic and distillation studies on rosemary (*Rosmarinus officinalis* L.) in a semi arid tropical environment. J. Herbs, Spices Med. Plants, 6(3): 25–30.
- PUANGSOMBAT K and SMITH J S (2010) Inhibition of heterocyclic amine formation in beef patties by ethanolic extracts of rosemary. J. Food Sci., **75**(2): T40–T47.
- RASHA, A R and ABDELLA, E M (2010) Modulatory effects of *rosemary* leaves aqueous extract on doxoruicin-Induced histological lesions, apoptosis and oxidative stress in mice, *Iranian* J. Cancer Preve., 3(1): 1–22.
- REZZOUG S A, BOUTEKEDJIRET C and ALAF K (2005) Optimization of operating conditions of rosemary essential oil extraction by a fast controlled pressure drop process using response surface methodology, J. Food Eng., **71**: 9–17
- RICHHEIMER S L, BERNART M W, KING G A, KENT M C and BAILEY D T (1996) Antioxidant activity of lipid soluble diterpenes from rosemary, J. Am. Oil Chem. Soc., **73**(4): 507–54.
- ROSE J (1974) Herbs and Things. Workman, New York.
- SAMMAN S, SANDSTROM B, TOFT M B, BUKHAVE K, JENSEN M, SORENSEN S S, HANSEN M (2001) Green tea or rosemary extract added to foods reduces nonheme-iron absorption, Am. J. Clin. Nutr., 73: 607–12.
- SANTOS GARCIA-ALVARDO J, JULIA VERDE-STAR M and HEREDIA N L (2001) Traditional uses and scientific knowledge of medicinal plants from Mexico and Central America, J. Herbs, Spices Med. Plants, 8(213): 37–90.
- SHAH S C, GUPTA L K, BHUJWAN H S and BHUJWAN S H (1996) Effect of auxins on rooting of stem cuttings of rosemary (*Rosmarinus officinalis* L.), *Recent Hort.*, **3**(1): 126–8.
- SILVA C DE P and PEDRAS J F (1999) Early rooting in rosemary (*Rosmarinus officinalis* L.) cuttings under the influence of chemical treatment and collecting time, *Acta Hort. (ISMS)*, **502**: 213–17.
- SINGH M and RAMESH S (2000) Effect of irrigation and nitrogen on herbage, oil yield and water use efficiency in rosemary grown under semi-arid tropical conditions, J. Med. Aromati. Plants Sci., 22(1B): 659–62.
- SINGH M, RAGHAVAN B and ABRAHAM K O (1996) Processing of marjoram (*Marjoram hortensis* M) and rosemary (*Rosmarinus officinalis* L) effect of blanching method on quality, *Nahrung*, 40(5): 264–6.
- SINGLETARY K, MACDONALD C and WALLIG M (1996) Inhibition by rosemary and carnosol of 7,12-dimethylbenz(a)anthracene (DMBA) induced rat mammary tumorigenesis and *in vitro* DMBA–DNA adduct formation, *Cancer Lett.*, **104**(1): 43–8.
- SOLINAS V, DEIANA S, GESSA C., BAZZONI A, LODDO M A and SATTA D (1996) Effect of water and nutritional conditions on the *Rosmarinus officinalis* L. – phenolic fraction and essential oil yields. *Seminario Internazionale di Chiusura del Progetto PIM, Sardegna, Cagliari*, 18–19 Nov. 1994, *Rivista Italiana EPPOS*, **19**: 189–98.
- TAKENAKA M, WATANABE T, SUGAHARA K, HARADA Y, YOSHIDA S and SUGAWARA F (1997) New antimicrobial substances against *Streptomyces scabies* from rosemary (*Rosmarinus officinalis* L.) leaves, *Biosci. Biotech. Biochem.*, **61**(9): 1440–44.
- TARRAF S and IBRAHIM M E (1999) Physiological response of rosemary (*Rosmarinus officinalis* L.) plant to brassinosteroid and uniconazole, *Egypt J. Hort.*, **26**(3): 405–19.
- TAWFIK A A (1997) In vitro selection for salt tolerance in rosemary (Rosmarinus officinalis L.), Assiut J. Agric. Sci., 28(4): 67–78.
- TENA M T, VALCARCEL M, HIDALGO P J and UBERA JL (1997) Supercritical fluid extraction of natural Rosemary oil does not detonateantioxidants from rosemary: comparison with liquid solvent sonication, *Analyt. Chem. Washington*, **69**(3): 521–6.

- VALENZUELA A B and NIETO S K (1996) Synthetic and natural antioxidants: food quality protectors, Grasas Y Acetias, 47: 186–96.
- VASUNDHARA M, KHAN M M, FAROOQI A A and NUTHAN D (2002) Effect of drip fertigation on yield and cost economics in rosemary (*R. officinalis* L.), *Indian Perfumer*, **46**(2): 179–90.
- VILLEVIEILLE M, CASSINI R and NICOT P (1999) Hook disease in tips of rosemary, *Phytoma*, **517**: 50–3.
- WEINBERG Z G, AKIRI B, POTOYEVSKI E and KANNER J (1999) Enhancement of polyphenol recovery from rosemary (*R. officinalis* L.) and sage (*Salvia officinalis* L.) by enzyme assisted ensiling (ENLAC), *J. Agric. Food. Chem.*, **47**(7): 2959–62.
- WOLSKI T, LUDWICZUKA A, ZWOLAN W and MARADAROWICZ M (2000) GC/MS analysis of the content and composition of essential oil in leaves and gallenic preparations from rosemary (*Rosmarinus officinalis* L.), *Herba Polonica*, 46(4): 243–8.
- YANG-RONG HUI, POTTER T P, CURTIS O F, SHETTY K and YANG R H (1997) Tissue culture based selection of high rosmarinic acid producing clones of rosemary (*Rosmarinus officinalis* L.) using *Pseudomonas* strain F, *Food. Biotechnol.*, **11**(1): 75–88.
- YANISHLIEVA-MASLAROVA and HEINONEN I M (2001) Rosemary and sage as antioxidants, in Peter K V (ed.), Handbook of Herbs and Spices, Vol. 1. Woodhead, Cambridge, 269–75.
- YESÏL-CELÏKTAS O, SEVÏMLÏ C, BEDÏR E and VARDAR-SUKAN F (2010) Inhibitory effects of rosemary extracts, carnosic acid and rosmarinic acid on the growth of various human cancer cell lines, *Plant Foods Hum. Nutri.*, **65**(2): 158-63.
- ZEGARSKA Z, AMAROWICZ R, KARMAC M and RAFLOWSKI R (1996) Anti oxidative effect of rosemary ethanolic extract on butter, *Milchwissenschaft*, **51**(4): 195–8.
- ZERMANE A, MENIAL A-H and BARTH D (2010) Supercritical CO<sub>2</sub> extraction of essential oil from Algerian rosemary (*R. officinalis* L.), *Chem. Eng. Technol.*, **33**: 489–98.
- ZHU-RU X, RAO H Y, GONG LAN, ZHU RX, RAO HY and GONG L (1996) Suspended cell culture of Rosmarinus officinalis and production of essential oil, Plant Physiol. Comm., 32(1): 9-12.