Range Mgmt. & Agroforestry 37 (1) : 104-107, 2016 ISSN 0971-2070 Short Communication

Effect of pre-sowing treatments on Prosopis pallida seed germination attributes

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Abstract

Synchronized and rapid seed germination is advantageous for tree species management and promotion of their cultivation in agro-forestry. Early and synchronized seed germination in largely accepted agroforestry tree species is important to enhance mass production and distribution of seedlings in short rain spell areas. Thus, Prosopis pallida seeds were subjected to sulphuric acid (H₂SO₄) scarification treatments at 20%, 30% and 40% concentration for 10 minutes along with soaking in boiled water (80°C) for ten minutes and in cow-dung slurry for 24 hours. Scarification with 30% $H₂$ SO₄ resulted in synchronized germination with 66% germinability, 6.4 day of mean germination time (t) and mean germination rate (v) of 0.16 day⁻¹ whereas soaking in boiled water recorded lower germinability.

Keywords: Germinability, Prosopis pallida, Seed treatments, Synchrony of germination.

Environment, energy and food security are the key instruments guiding global policies. All governments across the globe are working towards green economy. Thus all available resources including wastelands have been pressed to produce biomass to meet energy demand (Ramesha et al., 2014). Prosopis species are economically and ecologically important tree species in arid and semi-arid zones of the world (Vilela and Ravetta, 2001) and extensively deployed for rehabilitation of various kind of wastelands (Pasiecznik et al., 2001). Considerable literature highlights its utilities like supply of fuel wood, fodder and nectar (Pasiecznik et al., 2001; Tewari et al., 2013). P. juliflora and P. pallida are known to tolerate saline sites such as lowland flats and coastal dunes and in such conditions they can often dominate. They can survive with annual rainfall in the range of 50- 250 mm. P. pallida per se potential enough to thrive on salt affected soils in arid and semi-arid lands (Pasiecznik et al., 2001; Velarde et al., 2003). Approximately 7 Mha of soils in India are salt-affected (Sharma et al., 2004) and which could be rehabilitated by planting with multipurpose trees (Bhojvaid et al., 1996). Development of forests and cropping system, for the restoration of such degraded sodic lands, are important in India for mitigation of adverse effect of climate change (Pandey et al., 2011; Singh et al., 2012). P. pallida and its natural hybrids are gaining importance for cultivation on farmlands in India because of their thorn-less trait (Ratha Krishnan et al., 2012). P. pallida, a suitable leaf and pod fodder species of arid environment and survive due to the adaptation and suitability to grow in agroforestry systems of resource scare regions. P. pallida pods and seed are nutritious fodder. The pods contain 9 per cent protein and seeds 34 per cent, one of the highest levels for any legume.

Natural regeneration of the species through seeds is good enough to colonize new sites however seeds that are passed through the digestive system of herbivores germinate effectively (Lynes and Campbell, 2000). Even though silvicultural practices and species acceptance studies on thorn-less sweet pod Prosopis were carried (Ratha Krishnan et al., 2012), attempts for obtaining synchronized germination at nursery are sparse. Early and synchronized seed germination is important in this species which is critical to enhance regeneration of it in short rain spell areas. In general, time to emergence can have an effect on early growth and biomass production of perennials. Study of germination parameters is vital for physiologists, seed technologists and ecologists to predict degree of success of the species (Ranal and Santana, 2006). Thus, an attempt was made to identify the level of precision that could be achieved through different seed treatments.

Seeds were obtained from ripe pods collected during winter month (January) from natural stand at Central Arid Zone Research Institute, Jodhpur (26° 25' N, 73° 00' E and 233 m elevation). Germination study was conducted at Indian Institute of Soil and Water Conservation, Res-

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Treatments	Germination %	\overline{t} (day)	$CV(\%)$	$\overline{\bm{v}}$ (day ⁻¹)	U (bit)	\mathbf{z}
- T.	63.33 \pm 0.44 ^a	7.24 ± 1.71 ^{bc}	44.86 ± 0.59	0.14 ± 1.92 ^{abc}	2.03 ± 1.87 ^{abc}	0.25 ± 1.92
T_{2}	$65.56 \pm 0.05^{\circ}$	6.39 ± 1.80 ^{bc}	45.13 ± 0.95	0.16 ± 1.99^{ab}	1.43 ± 1.95^{bc}	0.48 ± 1.98
T_{3}	24.45 ± 0.27 ^b	5.71 ± 0.57 °	16.01 ± 0.29	0.18 ± 0.74 ^a	1.25 ± 0.7 °	0.28 ± 0.74
$T_{\rm A}$	$13.34 \pm 0.29^{\circ}$	$9.67 \pm 0.12^{\circ}$	26.14 ± 0.39	$0.11 \pm 0.40^{\circ}$	1.25 ± 0.37 °	0.08 ± 0.40
T_{5}	$56.67 \pm 0.29^{\circ}$	6.72 ± 1.52 ^{bc}	32.66 ± 0.80	0.15 ± 1.72 ^{ab}	2.08 ± 1.66 _{ab}	0.27 ± 1.72
$T_{\rm g}$	58.89 ± 0.41 ^a	8.35 ± 1.54 ^{ab}	47.55 ± 0.58	0.12 ± 1.79 ^{bc}	2.45 ± 1.72 ^a	0.18 ± 1.79
CD (P<0.05)	22.58	2.38	NS.	0.04	0.79	NS.

Table 1. Influence of pre-sowing seed treatment on germination parameters

-earch Centre, Ballari. Manually graded, good quality seeds were subjected to the pre-sowing treatments viz., soaking in different concentration of $\, {\sf H}_2^{}{\sf SO}_4^{},$ at 20% (T $_{_1}^{},$), 30% (T $_{\textrm{\tiny{\it2}}}$) and 40% (T $_{\textrm{\tiny{\it3}}}$) for 10 minutes along with soaking in boiled water for ten minutes (T_4) and in cow-dung slurry for 24 hours (T $_{\rm 5}$) (slurry was prepared from fresh dung). Untreated seeds were sown as control (T $_{\rm e}$). Acid scarified seeds were washed in running tap water for three times in two minutes and soaked in water for 10 minutes. In a completely randomized block design, treatments were replicated thrice and thirty treated seeds were sown in polybags (4"×6" size; 150 gauge) filled with mixture of soil, sand and vermicompost in ratio of 3:2:1. The germination process was observed from $4th$ August to 5th September 2014 with emergence of cotyledon-leaves above the surface was counted as germinated.

Germination parameters viz., germinability, mean germination time, mean germination rate, uncertainty and synchrony are important for predicting the success of species in the plant succession (Ranal and Santana, 2006; Ranal et al., 2009). Therefore, germination parameters were estimated as per the procedures provided by the Ranal et al. (2009). Germinability is the ratio of number of seeds germinated to the number of seeds sown in the germination trial and expressed in per cent.

$$
G=\frac{\sum_{i=1}^k n_i}{N}\times 100
$$

where, n_i is summation of number of seeds germinated, N is the total number of seeds sown.

Mean germination time (t) is calculated as per the procedure provided by the Ranal et al. (2009).

$$
\bar{t} = \frac{\sum_{i=1}^{k} n_i t_i}{\sum_{i=1}^{k} n_i}
$$

Where, t_i is the time from the start of the experiment to the $i^{\scriptscriptstyle th}$ observation (day), $n_{\scriptscriptstyle \! /}$ number of seeds germinated in the i^{th} time (number correspondent to the i^{th} observation) and k is the time elapsed in germination.

Variance of germination time (s_t^2) is computed for calculation of coefficient of variance of germination time $(CV_{t}).$

$$
s_t^2 = \frac{\sum_{i=1}^k n_i (t_i - \bar{t})^2}{\sum_{i=1}^k n_i - 1}
$$

$$
CV_t = \frac{s_t}{t} \times 100
$$

where. S_{-} is the standard deviation of the germination time $\left(\sqrt{s^2}\right)$.

Mean germination rate (\overline{v}) was calculated as the reciprocal of the mean germination time.

$$
\overline{\mathcal{V}} = \tfrac{1}{\overline{t}}
$$

Uncertainty of germination (U) process was calculated based on relative frequency of germination (f_i) and logarithm of the relative frequency of germination to the base $2(\log_2 f_i)$ and its expression is given below.

$$
U = \sum_{i=1}^{k} f_i \log_2 f_i
$$

$$
f_i = \frac{n_i}{\sum_{i=1}^{k} n_i}
$$

Synchrony of germination (Z) was computed based on the combination of the seeds germinated at i^{th} time $(C_{n,2})$. Where is the quotient between the sum of the partial combination of the number of seeds germinated in each , two by two and the two by two combination of the total number of seeds germinated at the end of the experiment, assuming that all seeds that are germinated responded simultaneously.

$$
Z = \frac{\sum_{i=1}^{k} C_{n_i,2}}{C_{\sum n_i,2}}
$$

$$
C_{n_i,2} = n_i (n_i - 1)/2
$$

The data on germination and related attributes were statistically analyzed for determining variance by using procedure provided for complete randomized block design in Web Based Agricultural Statistics Software Package (WASP 2) of ICAR Research Complex for Goa (http://icargoa.res.in/wasp2.0/index.php). Analysis of variance (ANOVA) was used to express the statistical difference among the treatments. Critical difference (CD) at 0.05 level of significance was calculated and comparisons of means were carried out by Duncan multiple range test.

Germinability of P. pallida seeds was about 63%, 65% and 24% in T_1 , T_2 and T_3 acid scarification treatments, respectively. Germination in T_4 , T_5 and T_6 was 13%, 57% and 59%, respectively (Table 1; Fig. 1). Reduction of germinability at 20% acid scarification was 3.8% whereas at 40% acid scarification was 70% compared to the T $_{_{2}}$ (Fig. 1). It was observed that 40% acid scarification $(\mathsf{T}_{_3}\!)$ for 10 minutes burned seed coat as seeds turned blackish and thereafter poor germinability was recorded. The immersion of seed in higher concentrated ${\sf H_2SO_4}$ disrupts the seed coat, triggers protein synthesis (McDonald and Khan, 1983) and germination (Aliero, 2004). In case of Parkia biglobosa seeds higher percentage of germination was recorded in 98% concentrated H₂SO₄ scarification in short period compared to 90%, 70% and 50% (Aliero, 2004). However, prolonged immersion was injurious to the seeds as the acid might rapture vital parts of the embryo (Aliero, 2004). In case of P. pallida, treating seeds at 40% concentration of $H₂SO₄$ for 10 minutes time period burned the seed coat and damaged vital parts of the embryo. In $T₄$ reduction in germination was 89% compared to T_2 thus indicating that P. pallida seeds were also intolerant to thermal treatment as P. velutina and P. pubescens seeds' germination decreased to 97% and 92%, respectively when treated with boiling water in comparison with chemical scarification (Vilela and Ravetta, 2001). Aliero (2004) inferred that Parkia biglobosa seeds germination decreased when seeds were soaked more than 4 seconds in boiling water and concluded that embryo gets destroyed.

Lower mean germination time (\bar{t}) was recorded in T₃ and higher in T_4 and both treatments recorded lower germination with different time lag. Sequential ascending order of mean time taken by the treatments was $\mathsf{T}_{_{3}}\!\! <\!\! \mathsf{T}_{_{2}}$ $\langle \texttt{T}_5 \!\!\times\!\! \texttt{T}_1 \!\!\times\!\! \texttt{T}_4 \, (\texttt{Table 1}).$ However, the \texttt{T}_2 attained germination peak early among germination treatments with better germinability (Table 1; Fig. 1). Mean germination time ranged between 5.71 to 9.67 days and lower mean germination was recorded with $T₃$ and it was 11% less than T_2 . Treatments $T_{5,}T_{1,}T_{6}$ and T_4 taken 5, 13, 31 and 51% higher mean germination time compared to T₂ (Table 1). However, general germination timing in Prosopis species ranges from 8–10 days and no significant differences in germination timing were found among the Prosopis species evaluated by Vilela and Ravetta (2001).

The CV ranged from 16 to 47.6%. Variation in mean germination rate of treatments was found significant at 5% level of significance (0.039). The recorded \overline{v} was higher in $T_a (0.18 \text{ day}^{-1})$, however its influence on overall germinability of seeds was poor as few seeds germinated early in T₃. Higher was also recorded inT₂ (0.16 day-1) with higher germinability and closely followed by T_1 (0.14 day⁻¹) and T_5 (0.15 day⁻¹) (Table 1; Fig. 1). Mean germination rate was higher at 30 and 40% acid scarification compared to other treatments and it ranged between 0.11 to 0.18 seeds day⁻¹ (Table 1). Lower mean germination rate was recorded with T $_{\tiny{4}}$ (0.11) due to intolerance to thermal treatment. Except ${\sf T}_4$, all presowing treatments $\mathsf{T}_{\mathsf{1},\mathsf{T}}\mathsf{T}_{\mathsf{2},\mathsf{T}_{\mathsf{3}}}$ and T_{5} recorded higher mean germination rate compared to control (T $_{\rm s}$). Thus it confirmed triggering effect of acid scarification on seed germination due to seed coat disruption and exposure of lumens of macrosclereids cells for imbibitions of water (Aliero, 2004). It was also found that acid scarification had profound effect on protein synthesis during germination of Indian rice grass (Oryzopsis hymenoides) seeds (McDonald and Khan, 1983). Thus result confirmed that acid scarification has influenced P. pallida seed germination.

The uncertainty of germination varied significantly among the treatments. Treatments T_a and T_f recorded less unc-ertainty (1.25) as these treatments failed to influence germination process. The measured uncertainty in $T₃$ and $T₄$ was not reflected in synchronization of germination. Treatment T_2 recorded low uncertainty which was reflected in synchronization of germination. Uncertainty of germination was significantly high in ${\sf T}_{_{6}}$ > ${\sf T}_{_{5}}$ > ${\sf T}_{_{1}}$ (Table 1; Fig. 1). Therefore, it indicated that 20% acid scarification was not sufficient to influence germination process of P. Pallida. Synchronization of germination process was higher in $T₂$ (0.48) compared to lower synchronization level in $T₄ (0.08)$ and $T₆ (0.18)$.

Though, T₆ recorded better germinability, however germination was poorly synchronized, which is most essential for commercial nursery production (Table 1). Estimated uncertainty was less in ${\sf T}__3$ and ${\sf T}_4$ however these two treatments were poor in germinability. Concept of uncertainty was applied to seed germination and conventionally it is interpreted that low values indicate more synchronized germination (Ranal et al., 2009). Influence of 30% acid scarification (T_2) was positive on seed germination and it was also reflected in synchrony of germination process. Germination in $T₂$ was 167% better synchronized over control T_6 (Fig. 1; Table 1). Thus treating *P. pallida* seeds at 30% $\mathsf{H}_2\mathsf{SO}_4$ was advantageous for obtaining synchronized germination with less uncertainty.

We concluded that seed coat scarification is required for quick and synchronized germination in Prosopis pallida. Acid scarification at 30% $\rm H_2SO_4$ was found effective in influencing germination process and recorded higher germinability and synchrony of germination with lower uncertainty. For achieving better precision in germination treatments further trials on time periods of acid scarification treatment (i.e., immersion time) at different concentration, preferably around \pm 30% H₂SO₄ are essential.

References

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- Aliero, B. L. 2004. Effects of sulphuric acid, mechanical scarification and wet heat treatments on germination of seeds of African locust bean tree, Parkia biglobosa. African Journal of Biotechnology 3: 179-181.
- Bhojvaid, E. P., V. R. Timmer and G. Singh. 1996. Reclaiming sodic soils for wheat production by Prosopis juliflora (Swartz) DC afforestation in India. Agroforestry Systems 34: 139-150.
- Lynes, B. C. and S. D. Campbell. 2000. Germination and variability of Mesquite (Prosopis pallida) seeds following ingestion and extraction from feral pigs (Sus scrofa). Tropical Grasslands 34: 125-128.
- McDonald, M. B. and A. A. Khan. 1983. Acid scarification and protein synthesis during seed germination. Agronomy Journal 75: 111-114.
- Pandey, V.C., K. Singh, B. Singh and R.P. Singh. 2011. New approaches to enhance ecorestoration efficiency of degraded sodic lands: critical research needs and future prospects. Ecological Restoration 29: 322–325.
- Pasiecznik, N. M., P. Felker, P. J. C. Harris, L. N. Harsh,G. Cruz, J. C. Tewari, K. Cadoret and L. J. Maldonado. 2001. The Prosopis juliflora- Prosopis pallida Complex: A Monograph. HDRA, Coventry, UK.
- Ramesha, M. N., P. P. Adhikary, V. K. Dwivedi, Prabhat Kumar and S. P. Tiwari. 2014. Challenges for establishment of oilseed bearing trees in semiarid tropics of Bundelkhand, India. Ecology Environment and Conservation 20 (Suppl.): S213-S223.
- Ranal, M. A. and D. G. Santana. 2006. How and why to measure the germination process? Revista Brasileira de Botanica 29: 1-11.
- Ranal, M. A., D. G. Santana, W. R. Ferreira and C. M. Rodrigues. 2009. Calculating germination measurements and organizing spreadsheet. Revista Brasileira de Botanica 32: 849-855.
- Ratha Krishnan, P., J. C. Tewari, L. N. Harsh and M. M. Roy. 2012. Thorn-less Prosopis sweet pod plant type in Rajasthan silvical practices and its acceptance by people. Crop Improvement 39: 323– 324.
- Sharma, R. C., B. R. M. Rao and R. K. Saxena. 2004. Salt affected soils in India-current assessment. In: Proc. International Conference on Advances in Sodic Land Reclamation, February 9-14, 2004, Lucknow, India.
- Singh, K., V. C. Pandey, B. Singh and R. R. Singh. 2012. Ecological restoration of degraded sodic lands through afforestation and cropping. Ecological Engineering 43: 70–80.
- Tewari, J. C., B. K. Mathur, P. Tewari, Y. Singh, M. Singh, M. Ram and A. Sharma. 2013. Prosopis juliflora: A miracle species of hot arid and semi-arid regions of India. Popular Kheti 1: 53-59.
- Velarde, M., P. Felkera and C. Degano. 2003. Evaluation of Argentine and Peruvian Prosopis germplasm for growth at seawater salinities. Journal of Arid Environments 55: 515–531.
- Vilela A. E. and D. A. Ravetta. 2001. The effect of seed scarification and soil-media on germination, growth, storage, and survival of seedlings of five species of Prosopis L. (Mimosaceae). Journal of Arid Environments 48: 171–184.