Determination of LD_{50} and LT_{50} of entomopathogenic nematodes against shoot borer (*Conogethes punctiferalis* Guen.) infesting ginger (*Zingiber officinale* Rosc.)

Rashid Pervez, S. Devasahayam and S.J. Eapen

Division of Crop Protection
Indian Institute of Spices Research, Kozhikode (Kerala) – 673 012, India

<rashid_pervez@rediffmail.com>

Date of Receipt: 10.01.2014; Accepted: 28.01.2014

ABSTRACT

Study on the pathogenicity of four native entomopathogenic nematodes, Heterorhabditis sp. (IISR-EPN 01), Steinernema sp. (IISR-EPN 02), Oscheius sp. (IISR-EPN 08) and O. gingeri against shoot borer larva (SBL), by dose response and time exposure assay and determination of lethal dosages (LD_{50}) and lethal time (LT_{50}) by regression analysis. LD_{50} were calculated using four densities of test EPNs viz., 25, 50, 75 and 100 IJs/SBL and LT_{50} was calculated using one nematode density 100 IJs to mortality of SBL at 24, 48 and 72 hrs. Although the SBL was susceptible to test EPNs, there were differences among these EPNs in their ability to kill the insect. Among test EPNs, Steinernema sp. (IISR-EPN 02) and O. gingeri appears to be the most promising. The per cent mortality of SBL increased with the increase the dosages as well as exposure time. A positive correlation was found between dosages as well as exposure time and mortality. Probit analysis indicated that Oscheius sp. (IISR-EPN 08) required less number (48 IJs/larva), whereas Steinernema sp. (IISR-EPN 02) took less time (29 hrs) for desired mortality of SBL. This study will be helpful in the shoot borer management under field conditions.

Key words: Conogethes punctiferalis, EPN, LD₅₀, Zingiber officinale.

Among the insect pests, shoot borer, Conogethes punctiferalis is one of the most important pest infesting Zingiberaceous crops specially ginger, turmeric and cardamom resulting in yield losses (Champakam et al., 2006 & Venugopal et al., 2006). The only effective method to manage this pest is the use of insecticides resulted residues in the produce affecting human health and also causing other ecological hazards. There has been a renewed interest in developing environment-friendly pest management schedules in agriculture. Entomopathogenic nematodes (EPNs) are effective biocontrol agents of a variety of economically important insect pests (Seal et al., 2010) and they have been used in controlling insect

pest for about a decade, extending their usage from high value markets to large area crops including spices (Pervez et al., 2012). Estimation of lethal concentrations and time are a relative measure of susceptible of insect and is convenient and commonly used index of relative efficacy (Girling et al., 2010). Hence, study on the pathogenicity of entomopathogenic nematodes, Heterorhabditis sp. (IISR-EPN 01), Steinernema sp. (IISR-EPN 02), Oscheius sp. (IISR-EPN 08) and O. gingeri (Pervez et al., 2013) against shoot borer larva, C. punctiferalis by dose response and time exposure assay and determination of lethal dosages and lethal time by regression analysis.

Materials and Methods

Infective juveniles (IJs) of test EPNs were obtained from stock cultures which maintained in the Nematology laboratory, I.I.S.R., Kozhikode. All EPN species were cultured on fully grown *Galleria mellonella* larvae as per the procedure described by Woodring and Kaya (1988). The IJs were surface sterilised in 0.1% hyamine solution and stored in distilled water in tissue culture flasks for study. *Galleria* larvae were obtained from cultures maintained on artificial diet. The shoot borer larvae were collected from ginger fields of the IISR-Experimental Farm, P'muzhi, Kozhikode. The larvae were sorted and those of similar size were select for the study.

Lethal dosages were calculated using four densities i.e. 25, 50, 75 and 100 IJs/SBL in 100 µl of distilled water. Suspension of each density was inoculated into six well plates (3.5 cm dia.) lined with filter paper at the bottom of the plate. To each well was introduced one shoot borer larva and the mortality of the larvae recorded at 72 hrs. The experiment was conducted at 28 °C in a BOD incubator and replicated 12 times along with controls. The mortality calculated into percentage and converted to probit mortality and values were transformed to log. LD₅₀ values were calculated using regression analysis. Lethal time was calculated using one density 100 IJs/SBL in 100 µl of distilled water. All experimental conditions were same as to determine LC₅₀ except the mortality of the larvae recorded at 24, 48 and 72 hrs of post inoculation and mortality calculated into percentage. The mortality was converted to probit mortality and exposure times were transformed to log values. LT₅₀ values were calculated using regression analysis.

Results and Discussion

All test EPNs were found pathogenic against SBL but the rate of mortality was vary. However, dosages and exposures time had a significant effect on mortality of the SBL. Per cent mortality of SBL increased with the increase the dosages of IJs. At 25 IJs/larva, mortality was first observed at 72 hrs for three test EPNs except *O. gingeri*, whereas it

was observed at 50 IJs/larva (Fig. 1). No larval mortality was recorded in control. A significant and positive correlation was found between dosages and mortality. Probit analysis indicated that, *Oscheius* sp. (IISR-EPN 08) and *Steinernema* sp. (IISR-EPN 02) required less number (48 & 50 IJs/larva, respectively) to provoke 50% mortality of SBL at 72 hrs post inoculation (Fig. 3). Bano *et al.* (2003) reported variable response of EPN species against red weevil.

The mortality induced by test EPNs was first observed at 24 hrs. SBL mortality % increased with the increase the exposure time (Fig. 2). A significant

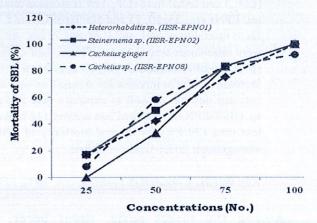


Fig. 1. Percent mortality of shoot borer larva (SBL) at different concentrations

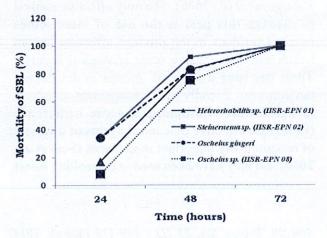


Fig. 2. Percent mortality of shoot borer larva (SBL) at different exposure time.

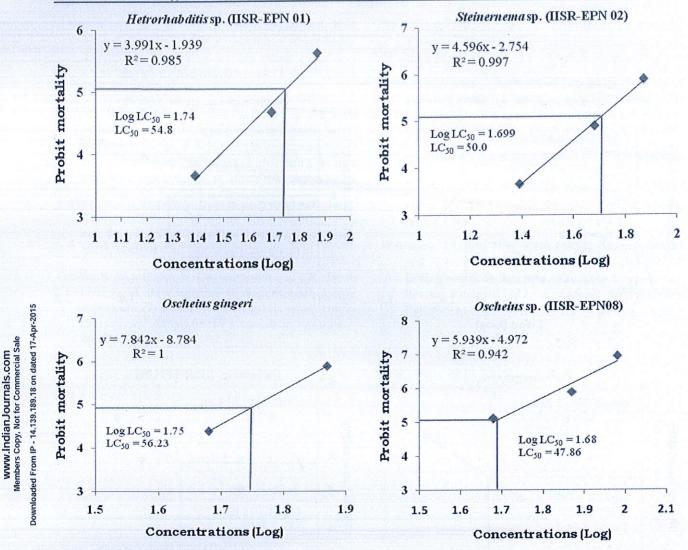


Fig. 3. LD₅₀ value of entomopathogenic nematodes against shoot borer larva.

and positive correlation was found between exposure time and mortality. Time assay response revealed that required *Steinernema* sp. (IISR-EPN 02) and *O. gingeri* took less time (29 & 31 hrs, respectively) to kill 50% SBL (Fig. 4). No larval mortality was observed in control. No statistical difference in mortality of SBL obtained among test EPNs at 100 IJs dosage. Therefore 25 and 50 IJs/larva was distinctive rates to differentiate EPNs biological efficacy on the shoot borer. Similarly, not much difference was recorded in the mortality of SBL among test EPNs at 72 hrs time exposure, whereas mortality at 24 hrs were distinctive rates

to differentiate infectivity of test EPNs against SBL. Saravanapriya and Subramanian (2007) reported dosage mortality response against stages of four lepidopterans insects. Although the shoot borer larva was susceptible to test EPNs, there were differences among these EPNs in their ability to kill the SBL. Among test of EPNs, *Steinernema* sp. (IISR-EPN 02), and *O. gingeri* appears to be the most promising EPN against SBL as it was reflected in the LD₅₀, LT₅₀ and % mortality. The mortalities were higher, LD₅₀ values were lower and LT₅₀ values were shorter for these EPNs. These differences may be due to difference of the origins

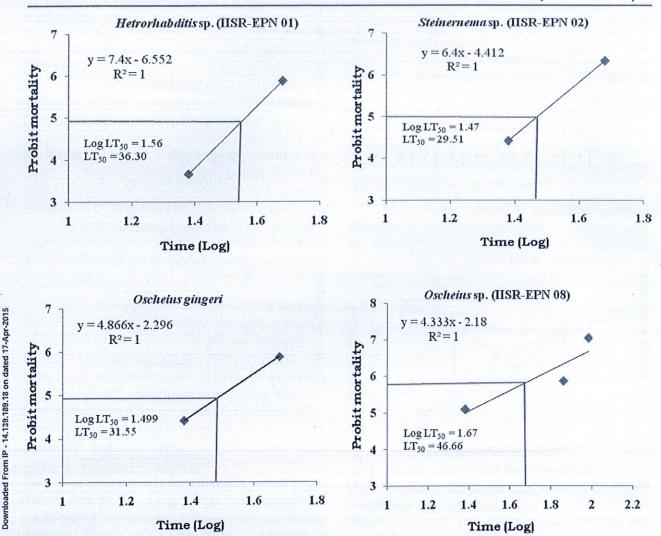


Fig. 4. LT₅₀ value of entomopathogenic nematodes against shoot borer larva.

of the strains (Canhilal, 2012). All strains of EPNs tested were effective in controlling *Agrotis ipsilon* and LD₅₀ was calculated to be 634 IJs (Seal *et al.*, 2010).

Apparently, they are effective biocontrol agents against shoot borer. However, environmental factors such as temperature, humidity and host density under green house and field conditions have huge impact on the efficacy of EPNs (Pervez & Ali, 2013). Therefore, future studies need to be directed to the green house and field conditions with these EPNs against shoot borer.

Acknowledgements: The authors express their gratitude to the Dr. M. Anandaraj, Director, Indian Institute of Spices Research, Kozhikode for providing all the facilities.

References

Bono, J. Gulsar, G. Rajendran and S.Subramanian (2003). Susceptibility of red weevil to entomopathogenic nematodes. *Ann. Pl. Protec. Sci.* 11: 104-106.

Canhilal, R. (2012). Comparison of efficacy of nine new heterorhabditid isolates (Rhabditida: Heterorhabditidae) in *Tenebrio molitor* (Coleoptera: Tenebrionidae). *African J. Microbio. Res.* 6: 1597-1602.

Members Copy, Not for Commercial Sale

www.IndianJournals.com

- Chempakam, B., R.S. Bhai, K. Kandiannan, K.S. Krishnamurthy and V.A. Parthasarathy (2006). Turmeric In: *Major spices-production and processing* (Eds.: V.A. Parthasarathy and P. Rajeev). I.I.S.R., Calicut, Kerala. pp. 152-190.
- Pervez, R. and S.S. Ali (2013). Entomopathogenic nematodes as a potent biopesticides against insect pests (Ed: Behra, K.K.), In: *Newer approaches to Biotechnology*, Narendra Publishing House Publishers and Distributors, New Delhi, pp. 213-235.
- Pervez, R., S.J. Eapen, S. Devasahayam and T.K. Jacob (2012). Efficacy of some entomopathogenic nematodes against insect pests of ginger and their multiplication. *Nematol. medit.* 40: 39-44.
- Pervez, R., S.J. Eapen, S. Devasahayam and T.K. Jacob (2013). A new species of entomopathogenic nematode *Oscheius gingeri* sp. n. from ginger rhizosphere. *Archieves of Phytopath. & Plant Prot.* 46: 526-535.

- Saravanapriya, B. and S. Subramanian (2007). Pathogenicity of EPN to certain foliar insect pests. *Ann. Pl. Protec. Sci.* **15**: 219-222.
- Seal, D.R., Vivek, K. Jha and T.X. Liu (2010). Potential of various strains of EPNs in combination with insecticides for suppression of black cutworm (Agrotis ipsilon). Ann. Pl. Protec. Sci. 18: 293-300.
- Venugopal, M.N., S.J. Ankegowda and D. Prasath (2006). Cardamom. In: *Major spices-production and processing*. (Eds. V.A. Parthasarathy and P. Rajeev). Indian Institute of Spices Research, Calicut, Kerala, pp. 62-114.
- Woodring, J.L. and H.K. Kaya (1988). Steinernematid and Heterorhabditid Nematodes: A Handbook of Biology and Techniques. Southern Cooperative Series Bulletin 331, Arkansas Agricultural Experiment Station, Arkansas, Fayetteville, 28 pp.