ISSN: 0367-973X

MULLI PHYTOPATHOLOGY

Well are

Signtember 1989

No. 3



(Issued January, 1990)

Published by
THE INDIAN PHYTOPATHOLOGICAL SOCIETY
(Indian Phytopath, 42(3): 369-485)

Effect of systemic fungicides on in-vitro growth of Pythium aphanidermatum, the rhizome rot pathogen of ginger

N. RAMACHANDRAN, G. N. DAKE AND Y. R. SARMA National Research Centre for Spices, Calicut 673 012

The rhizome rot of ginger (Zingiber officinale Rose.) caused by the fungus Pythium spp. is a major problem in all the ginger growing areas. Reports on in-vitro and in-vivo

efficacy of many conventional fungicides (1,9,11,12) on *P. aphanidermatum* are available, yet information with regard to the newer anti-pythiaceous systemic fungicides is either inadequate or lacking. We evaluated five chemicals representing four different groups of fuingicides for their efficacy on mycelial growth of the fungus.

The systemic fungicides used in this study included two phenylamides [metalaxyl technical 95.9 per cent, Hindustan Ciba-Geigy Ltd.; oxadixyl 25 W.P. Sandoz (India) Ltd.,]; one alkyl phosphonate [aliette 80 W.P. (fosetyl-Al) Rhone-Poulene Agrochemicals (India) Ltd.]; one carbamate [previcur-N 72.2 per cent (propamocarb) Schering AG] and ethazole [terrazole 35 W.P. (Olin Corporation)]. The stock solutions of the fungicides were prepared in sterile distilled water and then incorporated in required quantities to cornmeal agar medium before dispensing into petri plates. The concentration ranges tested were $0.01-10~\mu g/ml$ of metalaxyl and ethazole; $100-1000~\mu g/ml$ of fosetyl-Al and $0.01-100~\mu g/ml$ of oxadixyl and propamocarb. The water content of the cornmeal agar was so adjusted as to get a final volume of 50 ml for each concentration after adding the fungicide solution. This was uniformly distributed into three $100 \times 15~mm$ petri plates which served as replicates. Three plates containing unamended cornmeal agar were maintained as control.

P. aphanidermatum isolated from the collar regions of infected pseudostems of ginger and maintained in carrot agar slants was used in the studies. The petri plates were inoculated with 3 mm discs cut from advancing margins of a 3-day old culture grown on carrot agar. Because of the fast growth of the fungus the inoculum discs were kept towards the periphery of the plates in order to allow a longer incubation period. The inoculated plates were incubated at $25 \pm 1^{\circ}$ C for 48 h. The radial growth of the fungal colonies were measured as distance from the edge of the inoculum disc to the margins of the colonies extending through the centre of the plates. Linear regression analysis of the probit values of inhibition percentages and the log values of $100 \times 10^{\circ}$ concentrations was carried out to obtain the ED50 and ED90 values (Table 1).

TABLE 1: In-vitro effect of systemic fungicides on mycelial growth of Pythium aphanidermatum on cornmeal agar expressed as ED50 and ED90 (μg/ml).

Fungicides	ED50	ED90
Fosetyl-Al	293.70	934.10
Metalaxyl	0.74	9.50
Oxadixyl	27.18	179.60
Propamocarb	4.44	305.56
Ethazole	0.25	1.14

Ethazole was most toxic of all the fungicides tested and had the lowest ED‰ and ED‰ values (Table 1) followed by metalaxyl. The slope values of these fungicides were 1.94 and 1.15 respectively. Fosetyl-Al with a slope value of 2.55 had a low *in-vitro* toxicity against P. aphanidermatum. An indirect mode of action through modified host defence was attributed to fosetyl-Al earlier (2,7) but it is now reported to have a direct mode of action on the fungus (4,5,6,8). Propamocarb has a lower ED‰ (4.4 μ g/ml) but a higher ED‰ value (305.56 μ g/ml) compared to oxadixyl whose ED‰ and ED‰ values are 27.18 and 179.6 μ g/ml respectively. The correlation coefficients of the fungicides varied from 0.91 to 0.99. Though ethazole showed very high *in-vitro* toxicity, its low water solubility

may influence its systemicity in plants because water solubility is considered to be an important attribute of fungicides with selective action against comycetes which usually thrive in aqueous environments (3,10).

The authors are thankful to Mr. Jose Abraham for statistical analysis; the pesticide companies concerned for providing the fungicide samples; Dr. M. K. Nair, Director Central Plantation Crops Research Institute, Kasaragod, and Dr. A. Ramadasan, Joint Director, National Research Centre for Spices, Calicut, for providing facilities.

- 1. Abbaiah, K. and Subbayya, J. Andhra agric. J. 22 (3/4): 79-82 (1975).
- 2. Bompeix, G., Fettouche, P. and Saindrenan, P. Phytiatr. Phytopharm. 30: 257-272 (1981).
- Bruin, G. C. A. and Edgington, L. V. In Zoosporic Plant Pathogens—A Modern Perspective (Buczacki, S. T., Eds.), 193-232, Academic Press, London 352 pp (1983).
- 4. Dereks, W. and Buchenauer, H. Crop Prot. 6(2): 82-89 (1987).
- 5. Fenn, M. E. and Coffey, M. D. Phytopathology 74: 606-11 (1984).
- 6. Fenn, M. E. and Coffey, M. D. Phytopathology 75: 1064-68 (1985).
- 7. Guest, D. I. Physiol. Plant Pathol. 25: 125-34 (1984).
- 8. Jailloux, F., Bugaret, Y. and Froidefond, G. Crop. Prot. 6 (3): 148-152 (1987).
- Joshi, L. K. and Sharma, N. D. In Ginger and Turmeric (Nair, M. K., Premkumar, T., Ravindran, P. N. and Sarma, Y. R., Eds), 104-119, Central Plantation Crops Research Institute, Kasaragod (Kerala), India, 260 pp (1982).
- 10. Kerkenaar, A. and Kaars Sijpesteijn, A. Pestic. Biochem. Physiol. 15: 71-78 (1981).
- Sarma, Y. R., Nambiar, K. K. N. and Brahma, R. N. Proc. PLACROSYM-II, Indian Society for Plantation Crops, 386-397 CPCRI, Kasaragod (Kerala), India 555 pp (1979).
- 12. Sharma, N. D. and Joshi, L. K. Pesticides 13(8): 37-39 (1979).

Received for publication March 21, 1988.