NEW HOSTS OF CORYNESPORA CASSIICOLA (BERK. AND CURT.) WEI

Corynespora cassicola (Berk. and Curt.) Wei¹⁴ on brinjal and tomato has been occurring in Tirupati since 1965 during September-February period. During our studies on Corynespora leaf spot of brinjal, we noticed it on chilli, Solanum nigrum L., Ocimum sanctum L., Leucas aspera Spr., Croton sparsiflorus Mor, ¹² Gossypium hirsutum L., ³ Digera arvensis Forsk. and Syzygium jambolanum D.C. A comparative study of the disease in the above hosts is reported here.

Leaf spots on tomato and chilli are dark brown, 3-4 mm. in diameter and target type with a chlorotic halo. Stem infection as dark jrregular streaks is of common occurrence. Brinjal, Ocimum, Leucas and cotton show similar spotting but target type appearance is uncommon. In brinjal the lesions may coalesce to form irregular spots of 1 cm, size. Stem infection is not seen in these hosts. However dead stems of brinjal show velvety growth of the abundant conidiophores and conidia due to subsequent saprophytic colonization. Mature lesions on the above five hosts show abundant S. nigrum L., D. arvensis and sporulation. S. jambolanum develop minute, dark, nonsporulating spots. In the case of S. jambolanum, only seedlings at 6-8 leaf stage showed this type of infection and it was not noticed on trees.

Conidiophores are cylindrical, brownish, septate, with a bulbous basal cell. Conidia are obclavate to narrow cylindrical, in chains of 2 or 3, hyaline when young, later dark brown and thick-walled. They are porogenous and distoseptate, with a conspicuous hilum. Some abnormal bifurcate conidia have been noticed in the sporulating mass on dead stems of brinjal.

Isolates from all the hosts grow and sporulate well on potato-sucrose-agar (PSA). The range of the size of the conidia of the isolates in culture is $12\cdot8-326\cdot4~\mu~\times~6\cdot4-9\cdot6~\mu$ and 2-22 septate. Measurements of conidia and conidiophores from different hosts are given in Table I. The similarity of the conidia in culture is seen as against that of the natural materials which show greater variation.

Pathogenicity tests proved lack of host specificity among the 4 solanaceous isolates. Isolates of Ocimum showed pathogenicity both on Ocimum and brinjal. Isolates from Syzygium, Leucas and cotton were found to be

TABLE I

Data on conidiophores and conidia of isolates
of Corynespora cassicola (in microns)

Host	Conidia				Cenidiophores			
	Length	Width		Septa	Length	Width	Bu cell†	Septa
Chilli				_		•		
Max. Min.	275 · 2 35 · 2			23 2	$275 \cdot 2 \\ 95 \cdot 6$		16·0 9·6	10 3
Brinjal l Max. Min.	$432 \cdot 0$	22.8 4.8			630 · 0 96 · 0			13
Brinjal o Max. Min.	153-6			17 2	072 · 0 102 · 0			17 3
Tomato Max. Min.	140.8	22·4 3·2		36 3	384·0 54·0			
Ocimum Max. Min.	134.4				217·6 54·6			
Leucas a Max. Min.	spera S 138•0 30•6	22.8					-	01 8
<i>Gossypit</i> Max. Min.	um hirsi 147 24		$7 \cdot 5$	14 3	459 · 0 74 · 0	-		12 3

* Hilum: Diameter of the bilum. † Bu, cell: Bulbous basal cell of conidiophore.

pathogenic to brinjal but did not infect their respective hosts under our test conditions.

Irrespective of the source of the isolates, earliest symptoms were noticed in 5-7 days on chilli, S. nigrum L., O. sanctum L., and in 2 to 3 days on brinjal and tomato, the disease being severe on the latter two.

Out of the several hosts tried brinjal isolates were also pathogenic to soybean, sesame, Cajanus cajan Millisp. and Abelmoschus esculentus, Moench.

In India C. cassicola has been reported on brinjal, tomato, papaw, piper betle L., castor, rubber, eucalyptus, cassava, castor, rubber, eucalyptus, cassava, cassava, gendarussa L., Adathoda sp., Barleria cristata L., Carissa sp., Pycanthemum, Rauwolfia serpentina Benth, and Croton sparsiflorus Mor. It has been reported on chilliform Australia.

Thus chilli, S. nigrum L., O. sanctum L., L. aspera Spr., D. arvensis Forsk., S. jambolanum D.C. and Gossypium hirsutum, L. are new host records for India.

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- Mohanty, N. N. and Behera, B. C., Proc. Ind. Sci. Cong. Assoc., 1960, p. 330.
- 8. and Mahapatra, P. K., Indian Phytopath., 1968, 21, 275.
- 9. Munjal, R. L., and Gill, H. S., Ibid., 1962, 15, 269.
- 10. Roy, A. K., Ibid., 1965, 18, 327.
- Ramakrishnan, T. S., Proc. Ind. Acad. Sci., Sect. B, 1960, 51, 164.
- 12. Shome, S. K. and Mukharjee, N., Sci. & Cult., 1964, 30, 602.
- Tiromalachar, M. J. and Lacy, R. C., Sydewia, 1951, 5, 124.
- 14. Wei, C. T., Mycol, Pap., C.M.I., 1950, 34, 9.
- Wilson, K. I. and Rema Devi, L., Indian Phytopath., 1966, 19, 393.

Asha Ram and Lele, V. C., Indian Phytopath., 1968, 21, 347.

^{2.} Ellis, M. B., Mycol. Pap., C.M.I., 1957, 40, 65.

^{3.} Jones, J. P., Phytopathology, 1961, 51, 305.

^{4.} Karan, D., Mycopath, et. Mycol. appl., 1966, 30, 187.

Mohanty, U. N. and Mohanty, N, N., Sci. & Cult., 1955, 21, 330.

^{6.} Mohanty, N. N., Ibid., 1958, 23, 608.